

Taxonomic changes within Imatidiini and Hybosispini (Coleoptera: Chrysomelidae: Cassidinae)

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Abstract. Type specimens of various taxa of Imatidiini and Hybosispini were examined to verify their identity. The following taxonomic changes are proposed based on these comparisons. New combinations: *Cephaloleia apertura* (Staines, 2013) comb. nov. (from *Aslamidium* Borowiec, 1984), *Cephaloleia jataiensis* (Pic, 1923) comb. nov. (from *Xenispa* Baly, 1859), *Pseudimatidium bicoloricornis* (Pic, 1926) comb. nov. (from *Windsorispa* Sekerka, 2014), *Xanthispa miniacea* (Blanchard, 1843) comb. nov. (from *Homalispa* Baly, 1859), *Xenispa aeneipennis* (Baly, 1859) comb. nov. (from *Cephaloleia* Chevrolat, 1836), *Xenispa gilvipes* (Uhmman, 1930) comb. nov. (from *Cephaloleia*), *Hybosispa claripes* (Pic, 1923) comb. nov. (from *Solenispa* Weise, 1905), *Hybosispa delectabilis* (Staines, 1996) comb. nov. (from *Cephaloleia*), *Hybosispa sulciceps* (Baly, 1885) comb. nov. (from *Cephaloleia*), and *Hybosispa truncatipennis* (Baly, 1869) comb. nov. (from *Cephaloleia*). New synonyms: *Cephaloleia impressa* Uhmman, 1930 = *Demothispa clermonti* Pic, 1934 syn. nov.; *Xenispa atra* (Pic, 1926) = *Cephaloleia cyanea* Staines, 1996, syn. nov. Change in status: *Cephaloleia fernandoi* (Bondar, 1940) stat. restit. is removed from synonymy with *C. opaca* Baly, 1859. The monotypic genus *Serratispa* Staines, 2002 assign. nov. is transferred from Sceloenoplini to Imatidiini based on morphological characters. Three species are described: *Cephaloleia fouquei* sp. nov. from Bolivia and Peru, *C. renei* sp. nov. from Ecuador, and *Xenispa fouquei* sp. nov. from Venezuela. Keys to *Cephaloleia basalis* Weise, 1910 species-group, Venezuelan species of *Xenispa* Baly, 1859, and species of *Hybosispa* Weise, 1905 are proposed to simplify identification of respective taxa. Most of the taxa discussed herein are provided with colour photographs.

Key words. Coleoptera, Chrysomelidae, taxonomy, entomology, new species, new synonymy, new combination, keys to species, Neotropical Region

Introduction

Imatidiini is a large Neotropical tribe of cassidine beetles presently containing 401 described species classified in 22 genera and distributed from southern United States to northern Argentina. Species in this tribe are primarily associated with monocotyledonous plants, mainly belonging to Arecaceae, Bromeliaceae, Orchidaceae, Poaceae and nearly all families of Neotropical Zingiberales. Larval stadia are onisciform in shape and the final one enclosing the pupa. Adult beetles are characterized by usually smooth dorsum without costae on elytra and pronotum with a setigerous pore bearing a long seta situated in anterior corners or along anterior margin. I have recently revised the genera of Imatidiini and summarized all available information on morphology and biology of the tribe (SEKERKA 2014). The paper also included numerous taxonomic changes, mainly transfers from one genus to another due to wrong interpretation of the type species designation of *Demotispa* Baly, 1859. I verified most types of involved taxa, however, some placements remained tentative and based on primary descriptions as the respective type material was unavailable to me at that time. Since 2014 I had opportunity to examine most of the types not previously studied revealing additional misplaced taxa. The present contribution deals with these taxa and their proper placement.

I also reviewed types of several species of *Cephaloleia* Chevrolat, 1836 and found that they actually belong to Hybosispini. Originally this small tribe contained only five species placed in a single genus *Hybosispa* Weise, 1910. They are superficially very similar morphologically to Imatidiini but clearly differ in not bearing any setigerous pore on pronotum and having acute and more or less projecting inner margin of eye. With the transfers proposed here the tribe contains presently two genera and 18 species. Since the number of species in *Hybosispa* doubled due to new transfers I propose here a key to that genus.

Lastly, I describe three new, recently discovered species of Imatidiini. Two species were previously identified as *Cephaloleia kolbei* Weise, 1910 but clearly differ from the type. The third species is a high montane *Xenispa* Baly, 1859 from Venezuela, which is similar only to *X. pulchella* Baly, 1859, the type species of *Xenispa*. All three species are dedicated in loving memory to René Fouquè (1980–2016), who perished in a car accident in November 2016 and to whom this issue of *Acta Entomologica Musei Nationalis Pragae* is dedicated. We shall never forget René.

Material and methods

Taxonomic changes were made based upon study of respective type material. Type localities which were not problematic are interpreted; remaining ones are cited in original spelling with additional remarks in square brackets or plain if not requiring additional comments. Label data are cited for every examined type. All data are transcribed verbatim as they appeared on the respective labels. Double vertical bar (||) separates data on different labels and a single vertical bar (|) separates lines within each label. Additional comments are placed in square brackets. The following shortcuts are used to characterize individual labels: cb – cardboard label, g – green, hw – handwritten, p – printed, r – red, s – soft label, t – typed, y – yellow. All specimens of herein described new species are provided with following red and printed label:

‘HOLOTYPE [or PARATYPE respectively] | [Genus name] [♂ or ♀ symbol] | [species name] sp. nov. | L. Sekerka des. 2017’.

A Canon EOS 550D camera with Canon MP-E 65 mm lens at the aperture 5.4 were used to take numerous separate images at different focal planes and later composed for optimum focus using Helicon Focus software.

Specimens examined in this study are deposited in following collections:

BMNH	Natural History Museum, London, United Kingdom (Max Barclay & Michael Geiser);
BPBM	Bishop P. Bernice Museum, Honolulu, Hawaii, USA (Jim Boone);
CMNC	Canadian Museum of Nature, Ottawa, Canada (François Génier);
DSCC	Davide Sassi collection, Castelmarte, Italy;
DWPC	Donald Windsor collection, Ciudad de Panamá, Panama;
HNHM	Hungarian Natural History Museum, Budapest, Hungary (Ottó Merkl);
LSPC	Lukáš Sekerka collection, Prague, Czech Republic;
MCSNM	Museo Civico di Storia Naturale di Milano, Milan, Italy (Fabrizio Rigato);
MDVC	Mauro Daccordi collection, Verona, Italy;
MNHM	Muséum National d’Histoire Naturelle, Paris, France (Antoine Mantilleri);
MNKM	Museo de Historia Natural ‘Noel Kempff Mercado’, Santa Cruz de la Sierra, Bolivia (Julieta Ledezma);
MNRJ	Museu Nacional, Universidade Federal, Rio de Janeiro, Brazil (Marcela and Miguel Monné);
NHMW	Naturhistorisches Museum, Vienna, Austria (Harald Schilhammer);
SEMC	Snow Entomological Museum, University of Kansas, Lawrence, Kansas, USA (Michael Engel, Zack Falin);
SDEI	Senckenberg Deutsches Entomologisches Institut, Müncheberg, Germany (Konstantyn Nadein);
USNM	National Museum, Smithsonian Institution, Washington D.C., USA (Alexander Konstantinov).

Systematic part

Imatidiini Hope, 1840

Cephaloleia apertura (Staines, 2013) comb. nov.

(Fig. 1)

Aslamidium (*Neoaslamidium*) *apertura* Staines, 2013: 286 (original description incl. black-and-white photograph).

Type locality. Venezuela, Aragua State, Rancho Grande Biological Station, 1550 m a.s.l.

Type material examined. HOLOTYPE: glued, ‘VENEZUELA: Aragua | Rancho Grande Biol. Stn., 1550 m | 10°21’38”N, 67°41’38”W | 12-14 MAY 1998; J. Ashe, R. Brooks, P. Hanley | VEN1ABH98 029 ex: flight intercept trap [w, p, cb] || [bar code] | SM0128228 | KUNHM-ENT [w, p, cb] || HOLOTYPE | *Aslamidium* | (*Neoaslamidium*) | *apertura* Staines | des. C. L. Staines 2013 [r, p, cb]’ (SEMC).

Remarks. STAINES (2013) described this species based on a single specimen and placed it in the subgenus *Neoaslamidium* Borowiec, 1998 of the genus *Aslamidium* Borowiec, 1984 based on antennomere I being longer than antennomere II.

The genus *Aslamidium* is characterized by pronotum with distinctly explanate margin with conspicuous latero-basal impression on each side. The subgenus *Neoaslamidium* differs in pronotum being much narrower than the base of elytra and at least basally parallel-sided while the nominotypical subgenus has pronotum as broad as base of elytra and semicircular (SEKERKA 2014).

I studied the holotype of *A. apertura* (Fig. 1) and find it does not conform to the characteristics of the genus *Aslamidium*. It is true that species of *Aslamidium* have antennomere I

longer than II but this is not a unique diagnostic character for the genus as many other taxa among Imatidiini have this character. Moreover *A. apertura* also has antennomere III very long, somewhat shorter than antennomere I while all species of *Aslamidium* have antennomere I shorter than III. The present species has subquadratic pronotum with lateral margins very narrowly widened only in anterior half and lateral slopes of the disc without any impression; elytra with narrowly explanate margin and smooth outer edge. Based on above mentioned characters I find the species belongs to the genus *Cephaloleia* and I hereby transfer *A. apertura* to that genus.

Cephaloleia apertura has body size 4.3 mm, densely punctate vertex, head brown infusate with black, black pronotum with anterior 1/5 and lateral widened margin yellow, and elytra yellow with black diffused pattern forming 16 more or less isolated spots (Fig. 1). Such pattern is not present in any other described species as all similarly sized species have elytra either with only two to eight spots or vittae. *Cephaloleia apertura* also has antennomere I with short apical projection on lower side and is distinctly longer than III and the latter is twice as long as II while small species of *Cephaloleia* with yellow-black dorsum has antennomere I shorter than III and not expanded on apex.

The coordinates on the locality label do not accurately represent the collection site as they are at elevation of 930 m a.s.l. and situated in the Carabobo State. Therefore I do not mention them in the type locality.

Distribution. Venezuela: Aragua (STAINES 2013).

Cephaloleia atripes (Pic, 1926) comb. nov.

(Fig. 2)

Amplipalpa atripes Pic, 1926b: 8 (original description).

Type locality. Original type locality 'Brésil' specified by DESCARPENTRIES & VILLIERS (1959) to Brazil, Goiás, Rio Verde based on the label data of the holotype.

Type material examined. HOLOTYPE: glued, 'GOYAZ | RIO VERDE [w, p, cb] || coll Pic [w, hw by Pic, s] || n sp prés | caerulea [w, hw by Pic, s] || type [y, hw by Pic, s] || TYPE [r, p, cb] || atripes | n sp [w, hw by Pic, s]' (MNHN).

Remarks. PIC (1926b) described this species presumably based on a single specimen as the description is written in singular and only a single length measurement was given. Therefore I consider the specimen holotype by monotypy. Pic placed the species in the genus *Amplipalpa* Harold, 1875 (= *Oediopalpa* Baly, 1859) and compared it to *A. foveipennis* Pic, 1923. *Amplipalpa atripes* since its description has been listed only in catalogues (e.g. UHMANN 1957a, 1964).

I have studied the holotype and the species does not belong to the tribe Spilophorini as it has setigerous pores present only in anterior corners of pronotum. I find that it belongs to *Cephaloleia* as it fits with the concept of that genus (SEKERKA 2014). Among *Cephaloleia* it is very distinct and easy to separate because it has metallic blue elytra, serrate latero-apical margins of elytra, head abruptly constricted behind eyes and vertex separated from head by a deep sulcus thus the eyes appear to project laterally. There are only five uniformly metallic blue species of *Cephaloleia* (*C. caeruleata* Baly, 1875, *C. dilatata* Uhmman, 1948, *C. emarginata* Baly, 1875, *C. metallescens* Baly, 1885 and *C. tarsata* Baly, 1859) all differing in head continuously constricted behind eyes without sulcus separating vertex. *Cephaloleia atripes*

has also quite unusual shape of pronotum as it has anterior corners obliquely truncate (Fig. 2) while all uniformly metallic blue species of *Cephaloleia* have anterior corners rounded.

Distribution. Brazil: Goiás (DESCARPENTRIES & VILLIERS 1959).

***Cephaloleia fernandoi* (Bondar, 1940) stat. restit.**

(Fig. 3)

Himatidium fernandoi Bondar, 1940: 38 (original description).

Cephaloleia opaca (misidentifications): UHMANN (1968): 125 (partim, material from Bahia); STAINES & GARCÍA-ROBLEDO (2014): 239 (partim, material from Bahia).

Type locality. Brazil, Bahia, ‘municípios de Jequié e Nazareth’.

Type material examined. PARATYPES: 8 spec., glued in pairs and pinned on one pin ‘cotipo [r, s, hw] || Himatidium 2861 | fernandoi Bond. | n. sp. [w, hw, cb, bf] || 395 [w, hw, cb]’ (MNRJ); 1 spec., same data (LSPC); 1 spec., glued ‘Brasil | Bahia | Bondar [w, hw, cb] || Himatidium | fernandesi [sic!] | Bondar [w, hw by Monrós, cb] || cotipo [r, hw, s] || Himatidium | fernandoi | Bondar [w, hw, s] || Cephaloleia [hw] | opaca [hw] | Baly [hw] | F. Monrós det. 19[p]24[hw] [w, p + hw by Monrós, cb] || F. Monros | Collection | 1959 [w, p, cb] || NOT A TYPE | Described from one | specimen deposited | in the British Museum. | C.L. Staines 1987 [w, hw by Staines, s, red frame]’ (USNM); 4 spec., glued on separate cards pinned on one pin, ‘BRASIL | BAHIA | Bondar leg. [w, hw, cb] || F. Monros | Collection | 1959 [w, p, cb]’ (USNM).

Remarks. BONDAR (1940) described *H. fernandoi* based on a long series of specimens collected in Jequié and Nazareth in Brazilian state of Bahia. He collected the material on inflorescence of *Calathea ovata* (Nees & Mart.) Lindl. and *C. virginialis* Linden ex Regel. (Maranthaceae). Bondar did not specify the exact number of specimens but explicitly stated that the type is in his collection and cotypes in BMNH and MNRJ. I did not have the opportunity to study the collection of Bondar to see how he labelled the type. But since he explicitly mentioned type and cotypes I consider for now the specimens in MNRJ as paratypes. It is also unclear, which of the two municipalities is the actual type locality, therefore I cite both in the original spelling.

MONRÓS (1945) compared two paratypes of *H. fernandoi* (ex collection Bosq) with one specimen identified as *Cephaloleia opaca* Baly, 1859 by J. Weise (ex collection Bruch) and concluded that they belong to the same species thus he proposed the synonymy. Since that time *H. fernandoi* was listed as synonym of *C. opaca* (e.g. UHMANN 1957b; STAINES & GARCÍA-ROBLEDO 2014). UHMANN (1968) reported additional material of *C. opaca*: three specimens from Bahia collected by Bondar and three specimens from Santa Catarina. He supposed that specimens from Bahia could belong to *H. fernandoi* and noticed differences between the two populations, and suggested that more material should be studied to resolve these taxa. Finally, STAINES & GARCÍA-ROBLEDO (2014) published a redescription of *C. opaca*, which however, contains also characters of *C. fernandoi*. I reexamined their material and all records from Bahia belong to *C. fernandoi*. Moreover these specimens represent actual paratypes of that species. One of the USNM specimens bears red label ‘cotipo’. Based on additional label pinned under the specimen Staines suggested that it is not a type because the species [*C. opaca*] was described from a single specimen in BMNH. However, he misjudged the specimen as the cotype label is identical to MNRJ specimens and was most likely handwritten by Bondar, therefore it is surely part of the type series of *C. fernandoi*. I consider also the four remaining USNM specimens as paratypes as they were most likely pinned on one pin with the first specimen and provided only with the cotype label (like many additional paratypes

in MNRJ). Monrós certainly remounted the material but I have no doubts that all specimens belong to the original type series. See additional information under *C. opaca*.

Recently I had opportunity to study types of both taxa and without doubt *H. fernandoi* belongs to the genus *Cephaloleia*, however in my opinion it is distinct from *C. opaca* and thus I hereby restore its species rank. These two species form a distinct and unique species-group of *Cephaloleia* characterized by moderately large body (ca. 6 mm), distinctly impressed rows of punctures and elevated intervals while all other species of *Cephaloleia* lack markedly elevated intervals. *Cephaloleia fernandoi* can be easily separated by markedly thick antennae. See Table 1 for morphological differences among the two species.

Distribution. Brazil: Bahia (BONDAR 1940).

Table 1. Distinguishing characters between *C. fernandoi* (Bondar, 1940) and *C. opaca* Baly, 1859.

character / species	<i>Cephaloleia fernandoi</i> Fig. 3	<i>Cephaloleia opaca</i> Fig. 4
antennae	short and thick	long and slender
antennomeres III–XI	as wide as long	longer than wide
anterior corners of pronotum	obtuse and moderately projecting anterad	angulate and strongly projecting anterad
antero-lateral margin of pronotum	forming weak but distinct angle	forming weak angle, more distinct in males
lateral sides of pronotum	distinctly canaliculate	weakly canaliculate
length/width ratio of elytra	1.4	1.6
ventral colouration	yellow	thorax laterally black

Cephaloleia fouquei sp. nov.

(Fig. 9)

Cephaloleia kolbei (misidentifications): STAINES & GARCÍA-ROBLEDO (2014): 198 + Fig. 170 (partim, redescription, faunistics); JALINSKY et al. (2014): Figs 2C–D (noted); CHABOO & STAINES (2015): 389 (faunistics).

Type locality. Bolivia, Beni Department, Rurrenabaque.

Type material. HOLOTYPE: ♂, glued 'BOLIVIA Beni | Rurrenabaque | 18.-21.xi.1998 | V. Tichý lgt. [w, p, cb]' (NMPC). PARATYPES: **BOLIVIA:** BENI: 2 ♂♂ 3 ♀♀, same data as holotype (LSPC, 1 ♀ NMPC); 4 ♂♂ 4 ♀♀, 'BOLIVIA C.Tello | Beni | I-II-2000 | Rurrenabaque [w, p, cb]' (MCSNM); 4 ♂♂ 1 ♀, 'BOLIVIA C.Tello | Beni | VI-2000 | Rurrenabaque [w, p, cb]' (MCSNM, 1 ♂ LSPC); 17 ♂♂ 11 ♀♀, 'BOLIVIA C.Tello | Beni | VII-2000 | Rurrenabaque [w, p, cb]' (MCSNM, 3 ♂♂ 3 ♀♀ LSPC, 1 ♀ MNHN, 1 ♂ 1 ♀ SDEI); 1 ♂ 2 ♀♀, 'BOLIVIA C.Tello | Beni | IX-2000 | Rurrenabaque [w, p, cb]' (MCSNM); 9 ♂♂ 10 ♀♀, 'BOLIVIA | Rurrenabaque | VII-VIII-05 C.Tello [w, p, cb]' (MCSNM, 1 ♂ 1 ♀ BMNH, 1 ♂ 1 ♀ BPBM, 1 ♂ LSPC). **COCHABAMBA:** 1 ♂ 2 ♀♀, 'BOLIVIA | CHAPARE | Limbo 2000 m | I.952 | Coll. A. Martinez [w, hw, cb]' || F. Monros | Collection | 1959 [w, p, cb]' (USNM); 1 ♂, 'Bolivia | Chapare Limbo | 2000 m I-1952 | A. Martinez [w, hw, cb]' (USNM); 1 ♂, 'Bolivia | D° Cochabamba | p. Chapare | G. Roman 450 mts | Martinez leg. | IX.1953 [w, hw, cb, last row written vertically on left side] || ARGENTINA | 1968 Colln. | J. Daguerre [w, p, cb]' (USNM); 1 ♀, 'BOLIVIA, XII.1985, | Dto. Chapare, | Villa Cunari [probably erroneous and refers to Villa Tunari] [g, p, cb] || DUPLICATE | RETAINED | FROM THE | J. WIESNER | COLLECTION [w, p, cb] || Cephaloleia [hw] | kolbei Weise [hw] | det. C. L. Staines [w, p, cb, hw by Staines]'

(USNM). **PERU: Cuzco:** 1 ♀, 'Marcapata | Peru [w, p, cb] || Cephal. prope [hw] | kolbei Ws. [hw] | UHMANN DET.5[p]6[hw] [w, p + hw by Uhmman, cb]' (HNHM). **MADRE DE DIOS:** 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~500m E on | Tr.15 12.569211°S 70.100261°W | 272m 10.VI.2011 Jalinsky, Radocy, | Wertenberger ex.leaf roll Calathea | lutea PER-11-JJ-033 [w, p, cb] || [barcode] | SEMC0982970 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♀, same data as previous but 'SEMC0982974' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~500m E on | Tr.15 12.569211°S 70.100261°W | 272m 10.VI.2011 Jalinsky, Radocy, | Wertenberger ex.leaf roll Calathea | lutea PER-11-JJ-029 [w, p, cb] || [barcode] | SEMC0982891 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (LSPC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~500m E on | Tr.15 12.569211°S 70.100261°W | 272m 10.VI.2011 Jalinsky, Radocy, | Wertenberger ex.leaf roll Calathea | lutea PER-11-JJ-027 [w, p, cb] || [barcode] | SEMC0982882 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~500m E on | Tr.15 12.569211°S 70.100261°W | 272m 10.VI.2011 Jalinsky, Radocy, | Wertenberger ex.leaf roll Calathea | lutea PER-11-JJ-031 [w, p, cb] || [barcode] | SEMC0982900 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♂, same data as previous but 'SEMC0982902' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~500m E on | Tr.15 12.569211°S 70.100261°W | 272m 12.VI.2011 Jalinsky, Radocy, | Wertenberger ex.leaf roll Calathea | lutea PER-11-JJ-051 [w, p, cb] || [barcode] | SEMC0982983 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~100m to | intersctn Tr.13 12.569211°S | 70.100261°W 272m 13.VI.2011 | Jalinsky, Radocy, Wertenberger | ex.leaf roll Calathea lutea | PER-11-JJ-061 [w, p, cb] || [barcode] | SEMC0983056 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (LSPC); 1 ♀, same data as previous but 'SEMC0982999' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~100m to | intersctn Tr.13 12.569211°S | 70.100261°W 272m 13.VI.2011 | Jalinsky, Radocy, Wertenberger | ex.leaf roll Calathea lutea | PER-11-JJ-063 [w, p, cb] || [barcode] | SEMC0983075 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♂, same data as previous but 'SEMC0983018' (SEMC); 1 ♂, same data as previous but 'SEMC0983020' (SEMC); 1 ♂, same data as previous but 'SEMC0983021' (SEMC); 1 ♂, same data as previous but 'SEMC0983023' (USNM); 1 ♂, same data as previous but 'SEMC0983026' (LSPC); 1 ♂, same data as previous but 'SEMC0983028' (USNM); 1 ♀, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~100m to | intersctn Tr.13 12.569211°S | 70.100261°W 272m 13.VI.2011 | Jalinsky, Radocy, Wertenberger | ex.leaf roll Calathea lutea | PER-11-JJ-065 [w, p, cb] || [barcode] | SEMC0983053 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♀, same data as previous but 'SEMC0983054' (USNM); 1 ♀, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~100m to | intersctn Tr.13 12.569211°S | 70.100261°W 272m 13.VI.2011 | Jalinsky, Radocy, Wertenberger | ex.leaf roll Calathea lutea | PER-11-JJ-069 [w, p, cb] || [barcode] | SEMC0983060 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~100m to | intersctn Tr.13 12.569211°S | 70.100261°W 272m 13.VI.2011 | Jalinsky, Radocy, Wertenberger | ex.leaf roll Calathea lutea | PER-11-JJ-071 [w, p, cb] || [barcode] | SEMC0983065 | KUNHM-ENT [w, p, cb] || Cephaloleia kolbei | det. J. R. Jalinsky 2011 [w, p, cb] || Cephaloleia | kolbei Weise | det. C. L. Staines 2012 [w, p, cb]' (SEMC); 1 ♂, 'PERU: Madre de Dios Dept. | CICRA Field Station, ~100m to | intersctn Tr.13 12.569211°S | 70.100261°W 272m 14.VI.2011 | CS Chaboo, ex. sweeping | PER-11-CSC-029 [w, p, cb] || [barcode] | SEMC1021352 | KUNHM-ENT [w, p, cb] || Cephaloleia | kolbei | Weise 1910 | Det. ZHFalin '14 [w, p, cb]' (SEMC); 1 ♀, same data as previous but 'SEMC1021353' (SEMC); 1 ♀, same data as previous but 'SEMC1021355' (LSPC); 1 ♂, same data as previous but 'SEMC1021363' (LSPC); 1 ♂, same data as previous but 'SEMC1021364' (SEMC). **Pasco:** 1 ♂, 'PERU: Pasco Dept. | Villa Rica-Puerto Bermudas Rd. | 1350 m, 10°42'30"S, 75°10'6"W | 17 OCT

1999; R.Brooks | PERU1B99 051, ex: rolled *Heliconia* leaves [w, p, cb] || [barcode] | SM0142472 | KUNHM-ENT [w, p, cb] || Cephaloleia | kolbei Weise | det. C. Staines 2000 [w, p, cb]' (SEMC); 1 ♀, same data as previous but 'SM0142473' and without identification label (LSPC); 1 ♂, same data as previous but 'SM0142474' (LSPC); 1 ♂, same data as previous but 'SM0142475' (SEMC); 1 ♀, same data as previous but 'SM0142476' (SEMC); 1 ♂, same data as previous but 'SM0142477'; without identification label but with additional label 'DUPLICATE RETAINED | FROM THE UNIVERSITY | OF KANSAS | COLLECTION [w, p, cb]' (USNM). UCAYALI: 1 ♀, 'PERU: Ucayali Dept. | Tingo Maria-Pucallpa Rd. | Puente Chino, km 205 | 1300 m, 9°8'12"S, 75°47'20"W | 14 OCT 1999; R.Brooks | PERU1B99 007B | ex: rolled *Heliconia* leaves [w, p, cb] || [barcode] | SM0187453 | KUNHM-ENT [w, p, cb] || Cephaloleia | kolbei Weise | det. C. Staines 2012 [w, p, cb]' (SEMC); 1 ♀, same data as previous but 'SM0187454' and without det. label (LSPC); 1 ♀, same data as previous but 'SM0187455' (SEMC); 1 ♂, same data as previous but 'SM0187456' (SEMC); 1 ♂, same data as previous but 'SM0187457' (LSPC); 1 ♂, same data as previous but 'SM0187458' and without det. label (USNM).

Description. Body elongate, parallel-sided. Length 6.6–9.2 mm.

Body yellow, elytra with elongate black spot on each humerus and common transverse preapical black spot; apical 1/5 length of elytra yellow, lateral margins entirely yellow. Humeral spots always emarginate internally, preapical spot always emarginate centrally. Elytral pattern quite constant and only varies in depth of emargination of individual spots.

Frons with broad and low carina extending from clypeus and slightly projecting to vertex. Frons and vertex smooth, shiny, and moderately and sparsely punctate. Vertex flat and transverse, with moderately large and oval basal impression. Head moderately constricted beyond eyes. Eyes normal, large, not projecting from dorsal outline of head. Gena large, smooth, shiny and finely punctate. Mouthparts typical, not modified. Antennae moderately long, thick with two basal antennomeres sparsely pubescent, remainder densely pubescent. Antennomere I elongate, densely pubescent on ventral side and with apex slightly projecting anteriorly on ventral side. Antennomere II distinctly triangular in males and globose in females. Antennomeres III–IV elongate with distinctly projecting apices on inner side, thus appearing subtriangular; V–VI with very minutely projecting apices, remaining antennomeres unmodified. In females all antennomeres from III on unmodified with rounded apices. Length ratio of antennomeres: 100 : 54 : 59 : 52 : 54 : 41 : 51 : 45 : 54 : 56 : 87. Antennomere III 1.1× as long as II, antennomeres IV, V, VII, VIII, X subequal in length, VI and IX distinctly shorter. Antennomeres thicker from VIII onwards, VIII–XI subquadratic, 1.2× as long as wide. Antennomere XI 0.8× as long as I and with rounded apex.

Pronotum 1.1–1.2× as wide as long, subparallel-sided, more or less distinctly constricted around midlength and slightly diverging before anterior corners. Basal corners sharp and slightly divergent, thus pronotum appears slightly wider at base than around midlength. Anterior 1/6 regularly narrowing anteriorly and forming anterior corners. Anterior corners subacute, broad, and distinctly depressed thus separated from disc. Anterior margin deeply emarginate between corners, regularly convex. Setigerous pores not on tubercles, barely visible, situated anteriorly in curved points, and bearing single very long seta. Due to weakly delimited setigerous pores and quite fragile setae pronotum appears to lack setae in many paratypes. Lateral margins not explanate, slightly swollen and forming narrow and barely canaliculate ridge. Disc weakly and regularly convex without any impressions, shiny, moderately and sparsely macropunctate. Surface smooth with fine but quite visible shagreen. Basal margin bisinuate and smooth.

Scutellum subtriangular, smooth and distinctly less shiny than pronotum due to faint shagreen.

Base of elytra distinctly wider than base of pronotum. Basal margin smooth and bisinuate. Humeral angles broadly rounded and not projecting. Humeral calli distinctly convex, smooth, shiny and impunctate, not separated by explanate margin and thus forming anterior corners of elytra. Elytra overall regularly and weakly convex, smooth, moderately shiny with faint shagreen and without impressions. Punctuation moderately large, arranged in ten mostly regular and not impressed rows plus scutellar row, apical 1/5 of elytra (yellow portion between apical margin and transverse black spot) irregularly punctate. Scutellar row long, formed by ca. 14 punctures and reaching to basal 1/4 of elytra. Punctures in rows more or less regularly arranged, interspaces as wide as puncture diameter or narrower. Internal rows similar, penultimate row with larger punctures and ultimate with smaller and denser punctures. Ultimate row regular without distinct vacancies. Intervals smooth and shiny, 1–2× as wide as rows, not elevated or impressed. Interval 5 and mainly 7 with several additional irregular punctures. Lateral margins narrow, barely explanate and canaliculate in central part 3/5 of length. Explanate part smooth and shagreen, impunctate. Outer margin moderately swollen and smooth, in apical 1/5 declivous. Elytral apices rounded and slightly emarginate at suture.

Prosternal collar smooth and shiny. Prosternal process broad, strongly expanded apically and with weak medial impression. Epipleura smooth, shagreen, and bare. Thoracic ventrites finely and sparsely punctate, and shagreen. Abdominal ventrites densely micropunctate and pubescent with long semiadherent setae. Ventrites I and II fused, suture indicated only laterally.

Sexual dimorphism distinct, antennomeres II–III triangular in males and globose in females. Apical margin of ventrite V emarginate in males and bisinuate with slightly pronounced central part in females.

Legs normal, unmodified, last tarsomere elongate, projecting beyond sole of penultimate. Claws simple, divergent.

Variability. Specimens from lowlands always have antennomere I with long dense pubescence on underside, while specimens from mountains have short pubescence but otherwise are morphologically similar.

Differential diagnosis. *Cephaloleia fouquei* sp. nov. can be easily distinguished from the two similar species by densely pubescent ventral side of antennomere I while *C. renei* sp. nov. and *C. kolbei* Weise, 1910 have antennomere I only with fundamental short and adherent pubescence. *Cephaloleia kolbei* (Fig. 11) also differs in thick antennae with antennomeres II–IV strongly triangular and larger in size (above 10 mm). *Cephaloleia fouquei* has slender antennae with antennomeres II–III weakly triangular. *Cephaloleia renei* (Fig. 10) has similar size as *C. fouquei* but differs also (except antennomere I) in humeral spot on elytra always emarginate and antennomeres II–III more triangular. It also has coarser and denser punctuation of pronotum.

Host plant. Maranthaceae: *Calathea lutea* (Aubl.) E. Mey. ex Schult. (JALINSKY et al. 2014, as *C. kolbei*).

Etymology. The species is dedicated in loving memory to René Fouqu  (1980–2016), friend and enthusiastic entomologist, who was world specialist on the tenebrionid tribe Stenosini.

Remarks. STAINES & GARC A-ROBLEDO (2014) published numerous specimens from Bolivia,

Brazil, Ecuador and Peru having size 7–10 mm, yellow body with black humeral and common preapical spot on elytra, and attributed them to *C. kolbei*, which was until that time known only from Colombia. However, they did not examine the holotype. I did study the holotype and compared it to additional specimens with similar pattern as well as to the part of material published by STAINES & GARCÍA-ROBLEDO (2014). The holotype clearly differs from other populations published by STAINES & GARCÍA-ROBLEDO (2014) as it has very robust antennae with antennomeres II–IV strongly triangular while the other populations have antennae slender and antennomeres II–III barely triangular, basically only II has more or less triangular shape. *Cephaloleia kolbei* is also stouter and larger (length above 10.0 mm vs. 7.0–8.5 mm, exceptionally exceeding 9.0 mm). Further comparison of specimens revealed that the material belongs to two similar yet distinct species here described as *C. fouquei* sp. nov. and *C. renei* sp. nov. Therefore all subsequent records of *C. kolbei* belong to one of the newly described species and *C. kolbei* seems to be restricted to Pacific versant of the Cordillera Occidental in Colombia.

Records from Bolivia and Peru belong to *C. fouquei* sp. nov. Subsequently the same specimens from Peru were again published by JALINSKY et al. (2014) and CHABOO & STAINES (2015). The record from Brazil is based on a single specimen from São Paulo de Olivença deposited in USNM, which belongs to a different species group. The specimen is most similar to *C. alternans* Waterhouse, 1881 but not identical.

Distribution. Bolivia: Beni, Cochabamba and Peru: Cuzco, Madre de Dios, Pasco, Ucayali.

Cephaloleia impressa Uhmman, 1930

(Figs 5–6)

Cephaloleia impressa Uhmman, 1930a: 36 (original description).

Demothispa clermonti Pic, 1934: 2 (original description), **syn. nov.**

Type localities. *Cephaloleia impressa*: ‘Brasilien: Saõ Paulo’. *Demothispa clermonti*: ‘Brésil: Hansa’.

Type material examined. *Cephaloleia impressa*: HOLOTYPE: ♂, glued, ‘SAÕ PAULO | BRAS. MRÁZ LGT. | MUS. PRAGENSE [w, p, cb, bf] | Collect. | Spaeth [w, p, cb] || HOLOTYPE [r, t, s] || Cephaloleia | impressa n. sp. | Det. Uhmman 28. [w, hw by Uhmman, cb]’ (NHMW). *Demothispa clermonti*: HOLOTYPE: ♂, glued, ‘Hansa | S.Catharina-Brésil | Coll.J.Clermont [w, p, s] || type [y, hw by Pic, s] || TYPE [r, p, cb] || Clermonti | n sp [w, hw by Pic, s]’ (MNHN).

Remarks. Recently I had opportunity to compare types of *Cephaloleia impressa* (Fig. 5) and *Demothispa clermonti* (Fig. 6) and found that they are conspecific. The holotype of *D. clermonti* has slightly paler colouration thus its punctuation appears coarser while the holotype of *C. impressa* has elytra deep pitch black and thus the punctuation appears somewhat finer. Otherwise the two specimens have identical shape, size and proportions therefore I synonymize *D. clermonti* with *C. impressa*.

Cephaloleia impressa belongs to a peculiar species-group with two more species – *C. basalis* (Weise, 1910) and *C. jataiensis* (Pic, 1923). All three share small size of body (length 3.5–4.1 mm), quite convex elytra with lateral constriction in the basal 1/3 of the disc, and minutely but distinctly serrate apico-lateral margins of elytra (at magnification 40×). *Cephaloleia basalis* and *C. impressa* share the same short shape and convexity of body while *C. jataiensis* is less convex and more elongate. All three species can be distinguished using the key below *C. jataiensis*.

Distribution. Brazil: Santa Catarina (Pic 1934) and Saõ Paulo (UHMANN 1930a).

***Cephaloleia jataiensis* (Pic, 1923) comb. nov.**

(Fig. 7)

Demothispa jataiensis Pic, 1923: 8 (original description).**Type locality.** Original type locality 'Brésil' specified by DESCARPENTRIES & VILLIERS (1959) to Brazil, Goiás, Jatahy based on the label data of the holotype.**Type material examined.** HOLOTYPE: ♀, glued, 'Jatahy | Prov.Goyas.Brésil [g, p, cb] || TYPE [w, p in red, cb] || Demothispa | jatahensis | m. [w, hw by Weise, s] || TYPE [w, p in red, cb] || jataiensis Pic | (1923) [w, hw by Pic, s]' (MNHN).**Remarks.** I have tentatively placed this species to *Xenispa* Baly, 1859 in my previous paper (SEKERKA 2014) because of very finely serrate apical margin of elytra and curious shape of body, however I was not entirely convinced and suggested it might belong also to *Cephaloleia*. Recently I had opportunity to compare the holotype of *D. jataiensis* with those of *D. clermonti* and *C. impressa* and found that they belong to the same species-group together with *C. basalis* (Weise, 1910), thus *D. jataiensis* is here transferred to *Cephaloleia*. The species can be separated using a key given below. For further comments see also remarks under *C. impressa*.**Distribution.** Brazil: Goiás (DESCARPENTRIES & VILLIERS 1959).**Key to *Cephaloleia basalis* species-group**

- 1 Anterior corners of pronotum strongly projecting anteriad. Antennomere I simple. Vertex sparsely and moderately densely punctate, not impressed. Elytra strongly convex, lateral impressions on disc deep. 2
- Anterior corners of pronotum barely projecting anteriad. Antennomere I with a small acute tip projecting anteriad on ventral side. Vertex coarsely and densely punctate, distinctly impressed. Elytra moderately convex, lateral impressions on disc shallow. *C. jataiensis* (Pic, 1923)
- 2 Base of elytra with large common red patch. Pronotum narrower, 1.3× as wide as long. Vertex strongly shiny, only with macropunctuation and micropunctuation. *C. basalis* (Weise, 1910)
- Elytra uniformly pitch black. Pronotum broader, 1.5× as wide as long. Vertex somewhat opaque due to microreticulation. *C. impressa* Uhmann, 1930

***Cephaloleia opaca* Baly, 1859**

(Fig. 4)

Cephaloleia opaca Baly, 1859: 62 (original description). UHMANN (1968): 125 (partim, material from Santa Catarina); STAINES & GARCÍA-ROBLEDO (2014): 239 (partim, material from Rio de Janeiro and Santa Catarina).**Type locality.** 'Brazil'.**Type material examined.** HOLOTYPE: pinned, 'Type | H. T. [w, p, s, circle label with red frame] || Baly Coll. [w, p, cb] || Cephaloleia | opaca | Baly | Brazil [g, hw by Baly, cb]' (BMNH).**Additional material examined.** BRAZIL: RIO DE JANEIRO: Rio de Janeiro, 1883, 1 spec., P. Germain lgt. (USNM), 1945, 1 spec., Dr. Nick lgt. (USNM); Trinidad near Parati, 2.–10.xii.2000, 2 spec., A. Kudrna jr. lgt. (LSPC). SANTA CATARINA: Blumenau, 1 spec. (USNM); Corupá ([formerly] Hansa Humboldt), i.1947, 2 spec. (USNM, LSPC).**Remarks.** BALY (1859) described *C. opaca* explicitly from a single specimen. UHMANN (1968) and STAINES & GARCÍA-ROBLEDO (2014) published additional records from Bahia,

Rio de Janeiro, Santa Catarina, Peru and Venezuela. The material from Bahia belongs to *C. fernandoi* (see further Remarks under that species). Records from Rio de Janeiro and Santa Catarina belong to *C. opaca* and the material from Santa Catarina represents the dark form mentioned by UHMANN (1968). Specimens from Venezuela belong to *Pseudostilpnaspis* cf. *curvipes* (Uhmann, 1951). I did not examine the material from Peru but I question this record as *C. opaca* seems to be restricted to lowlands along the Brazilian Atlantic coast. Therefore I do not include Peru and Venezuela in distribution of *C. opaca*.

The material belonging to *C. opaca* published by STAINES & GARCÍA-ROBLEDO (2014) is repeated here as the authors did not state number of specimens, collectors, etc. The specimen from Blumenau was published as the record from Brazil without further data. The specimen collected by P. Germain was figured (STAINES & GARCÍA-ROBLEDO 2014: Fig. 200).

Distribution. Brazil: Rio de Janeiro and Santa Catarina (UHMANN 1968, STAINES & GARCÍA-ROBLEDO 2014, present paper).

Cephaloleia renei sp. nov.

(Fig. 10)

Cephaloleia kolbei (misidentification): STAINES & GARCÍA-ROBLEDO (2014): 198 (partim – Ecuadorian specimens).

Type locality. Ecuador, Napo Province, 2 km S of Puerto Misahuallí, 01°02'46"S, 77°39'23"W, 450 m a.s.l.

Type material. HOLOTYPE: ♂, glued, 'ECUADOR Napo prov. | 2 km S of Puerto Misahuallí | 01°02'46"S, 77°39'23"W, 450m | second growth 8.xii.2009 | MAR: Calathea sp. | L. Sekerka & K. Štajerová lgt. [g, p, cb]' (NMPC). PARATYPES: **ECUADOR: Napo:** 13 ♂♂ 9 ♀♀, same data as holotype (LSPC, 1 ♀ NMPC); 2 ♂♂ 4 ♀♀, 'ECUADOR Napo prov. | 2 km S of Puerto Misahuallí | 439m, 01°02'S / 77°39'W, 450m | 8 XII 2009 | leg. L. Borowiec [w, p, cb]' (1 ♂ 1 ♀ BMNH, 2 ♀♀ MCSNM, 1 ♂ 1 ♀ SDEI); 4 ♂♂ 8 ♀♀, 'ECUADOR Napo prov. | Río Puno (10 km SE of | Puerto Misahuallí), 417m | 01°02'S / 77°36'W | 13 XII 2009 leg. L. Borowiec [w, p, cb]' (LSPC, 1 ♂ 1 ♀ MNHN, 1 ♂ 1 ♀ SEMC); 2 ♂♂ 1 ♀, glued, 'ECUADOR Napo prov. | Río Hollín 6.xii.2009 | Narupa-Loreto Rd. 1068m | 0°43'04"S, 77°38'19"W | MAR: Calathea sp. | L. Sekerka & K. Štajerová lgt. [g, p, cb]' (LSPC). **ORELLANA:** 1 ♀, glued, 'ECUADOR: Napo | Huaticocha | 0°45'22'S/77°27.84'W | VIII-19-1997 | Fred G. Andrews [w, p, cb] || DUPLICATE | RETAINED FROM | THE CALF. DEPT. | FOOD & AGRIC. | COLLECTION [w, p, cb]' (USNM); 1 ♂, 'ECUADOR Orellana prov. | Río Suyuno (7 km N of Loreto) | 426m, | 00°37'S / 77°17'W, | 10 XII 2009, leg. L. Borowiec [w, p, cb]' (LSPC). **SUCUMBÍOS:** 2 ♂♂ 2 ♀♀, 4 spec., glued, 'ECUADOR Sucumbíos pr. | Cascada San Rafael | 00°06'15"S, 77°35'13"W | montane forest 1300m | MAR: Calathea sp. | D. Windsor lgt. 20.-25.vii.2008 [g, p, cb]' (1 ♂ 1 ♀ BPBM, 1 ♂ 1 ♀ LSPC, 4 spec. DWPC); 1 ♀, glued, 'ECUADOR: Napo | Limoncocha | 3 June 1977 | W. E. Steiner [w, p, cb] || *m [w, p, cb]' (USNM).

Description. Body elongate, parallel-sided. Length: 7.6–9.5 mm.

Body yellow, elytra with elongate black spot on each humerus and common transverse preapical black spot; apical 1/5 length of elytra yellow, lateral margins entirely yellow. Humeral spots not obviously emarginate, slightly vary in size. Transverse preapical spot slightly to distinctly emarginate on anterior margin.

Frons with broad and low carina extending from clypeus but not continuing to vertex. Frons and vertex smooth, shiny, and finely and sparsely punctate. Vertex flat and transverse, with moderately deep circular basal impression. Head moderately constricted beyond eyes. Eyes normal, large, not projecting from dorsal outline of head. Gena large, smooth, shiny and finely punctate. Mouthparts typical, not modified. Antennae moderately long, thick with two basal antennomeres sparsely pubescent, remainder densely pubescent. Antennomere I elongate, only with short and sparse pubescence and with apex slightly projecting anteriorly on ventral

side. Antennomere II globose in both sexes. Antennomeres III–IV elongate with distinctly projecting apices on inner side, thus appearing subtriangular; V–VI with very minutely projecting apices, remaining antennomeres unmodified. In females all antennomeres from III on unmodified with rounded apices. Length ratio of antennomeres: 100 : 69 : 75 : 68 : 66 : 62 : 62 : 65 : 69 : 69 : 112. Antennomere III 1.1× as long as II, antennomeres IV, V, VIII, IX, X subequal in length, VI and VII equally long. Antennomeres thicker from VI onwards, VII–XI subquadrate, 1.2× as long as wide. Antennomere XI 1.1× as long as I and with rounded apex.

Pronotum as wide as long, for most of length parallel-sided, basal corners sharp and slightly divergent, thus pronotum appears slightly wider at base than around midlength. Anterior 1/6 regularly narrowing anteriorly and forming anterior corners. Anterior corners subacute, broad, and distinctly depressed thus separated from disc. Anterior margin deeply emarginate between corners, regularly convex. Setigerous pores not on tubercles, barely visible, situated anteriorly in curved points, and bearing single very long seta. Due to weakly delimited setigerous pores and quite fragile setae pronotum appears to lack setae in most of paratypes. Lateral margins not explanate, slightly swollen and forming narrow and barely canaliculate ridge. Disc weakly and regularly convex without any impressions, strongly shiny, finely and sparsely macropunctate. Surface smooth with faint and barely visible shagreen. Basal margin bisinuate and smooth.

Scutellum subtriangular, smooth and distinctly less shiny than pronotum due to faint shagreen.

Base of elytra distinctly wider than base of pronotum. Basal margin smooth and bisinuate. Humeral angles broadly rounded and not projecting. Humeral calli distinctly convex, smooth, shiny and impunctate, not separated by explanate margin and thus forming anterior corners of elytra. Elytra overall regularly and weakly convex, smooth, moderately shiny with faint shagreen and without impressions. Punctuation moderately large, arranged in ten mostly regular and not impressed rows plus scutellar row, apical 1/5 of elytra (yellow portion between apical margin and transverse black spot) irregularly punctate. Scutellar row long, formed by ca. 14 punctures and reaching to basal 1/4 of elytra. Punctures in rows more or less regularly arranged, interspaces as wide as puncture diameter or narrower. First and ultimate rows with smaller and denser arranged punctures than remaining rows. Ultimate row regular without distinct vacancies. Intervals smooth and shiny, 1–2× as wide as rows, not elevated or impressed. Interval 5 and mainly 7 with several additional irregular punctures. Lateral margins narrow, barely explanate and canaliculate in central part 3/5 of length. Explanate part smooth and shagreen, impunctate. Outer margin moderately swollen and smooth, in apical 1/5 declivous. Elytral apices rounded and slightly emarginate at suture.

Prosternal collar smooth and shiny. Prosternal process broad, strongly expanded apically and with weak medial impression. Epipleura smooth, shagreen, and bare. Thoracic ventrites finely and sparsely punctate, and shagreen. Abdominal ventrites densely micropunctate and pubescent with long semiadherent setae. Ventrites I and II fused, suture indicated only laterally.

Sexual dimorphism more or less distinct, antennomeres III–IV with slight projection on inner side of apex in males and globose in females. Apical margin of ventrite V emarginate in males and bisinuate with slightly pronounced central part in females.

Legs normal, unmodified, last tarsomere elongate, projecting beyond sole of penultimate. Claws simple, divergent.

Differential diagnosis. *Cephaloleia renei* sp. nov. is most similar to *C. fouquei* sp. nov. (Fig. 9) as both have similar size and shape. The latter differs in antennomere I with dense and long pubescence on ventral side (vs. without such pubescence), emarginate humeral spot on elytra (vs. not emarginate), antennomeres II–III weakly but distinctly triangular (vs. barely triangular), and pronotum moderately densely punctate with moderately coarse punctures (vs. sparsely punctate with much finer punctures). *Cephaloleia kolbei* (Fig. 11) differs in larger size, length above 10 mm (vs. 7.0–8.5 mm) and strongly triangular antennomeres II–IV (vs. only II–III barely triangular). See also differential diagnosis under *C. fouquei*.

Host plant. Maranthaceae: *Calathea variegata* (K. Koch) Linden ex Körn.

Biology. Specimens from the type locality were collected inside young rolled leaves of *Calathea variegata*. Remaining specimens were collected also in leaf-rolls of various species of *Calathea*. Adult beetles as well as larvae were clearly eating leaf surface when the leaf was unrolled.

Etymology. The species is dedicated in loving memory to René Fouqu  (1980–2016), friend and enthusiastic entomologist, who was world specialist on the tenebrionid tribe Stenosini.

Remarks. Specimens of *C. kolbei* without locality data and those from Ecuador (all in USNM) published by STAINES & GARC A-ROBLEDO (2014) belong to *C. renei* sp. nov. The three specimens without locality labels are strongly damaged and thus are not included in the type series. The two from Ecuador are designated as paratypes. See also remarks under *C. fouquei* sp. nov.

Distribution. Ecuador: Napo, Orellana and Sucumb os.

Pseudimatidium bicoloricornis (Pic, 1926) comb. nov.

Demothispa bicoloricornis Pic, 1926a: 14 (original description).

Type locality. Original type locality ‘Guyane Fr.’ specified by DESCARPENTRIES & VILLIERS (1959) to French Guyana: Saint-Laurent-du-Maroni based on the label data of the holotype.

Type material examined. HOLOTYPE: glued, ‘GUYANE FRAN  SE | St-Jean du Maroni | COLLECTION LE MOULT [w, p, cb] || coll Pic [w, hw by Pic, s] || type [y, hw by Pic, s] || TYPE [r, p, cb] || Museum Paris | Coll. M. Pic [b, p, cb] || bicoloricornis | n sp [w, hw by Pic, s]’ (MNHN).

Remarks. When I was assembling the data for my previous paper on Imatidiini (SEKERKA 2014) I tentatively placed this species to *Windsorisa* Sekerka, 2014 because PIC (1926a) compared it to *W. latifrons* (Weise, 1910) and the type was on loan to another researcher. Last year I had opportunity to study the holotype and found that it in fact belongs to the genus *Pseudimatidium* Aslam, 1966 and is most similar to *P. madoni* (Pic, 1936), which differs in stronger punctuation of elytra and pronotum.

Distribution. French Guyana: Saint-Laurent-du-Maroni (DESCARPENTRIES & VILLIERS 1959).

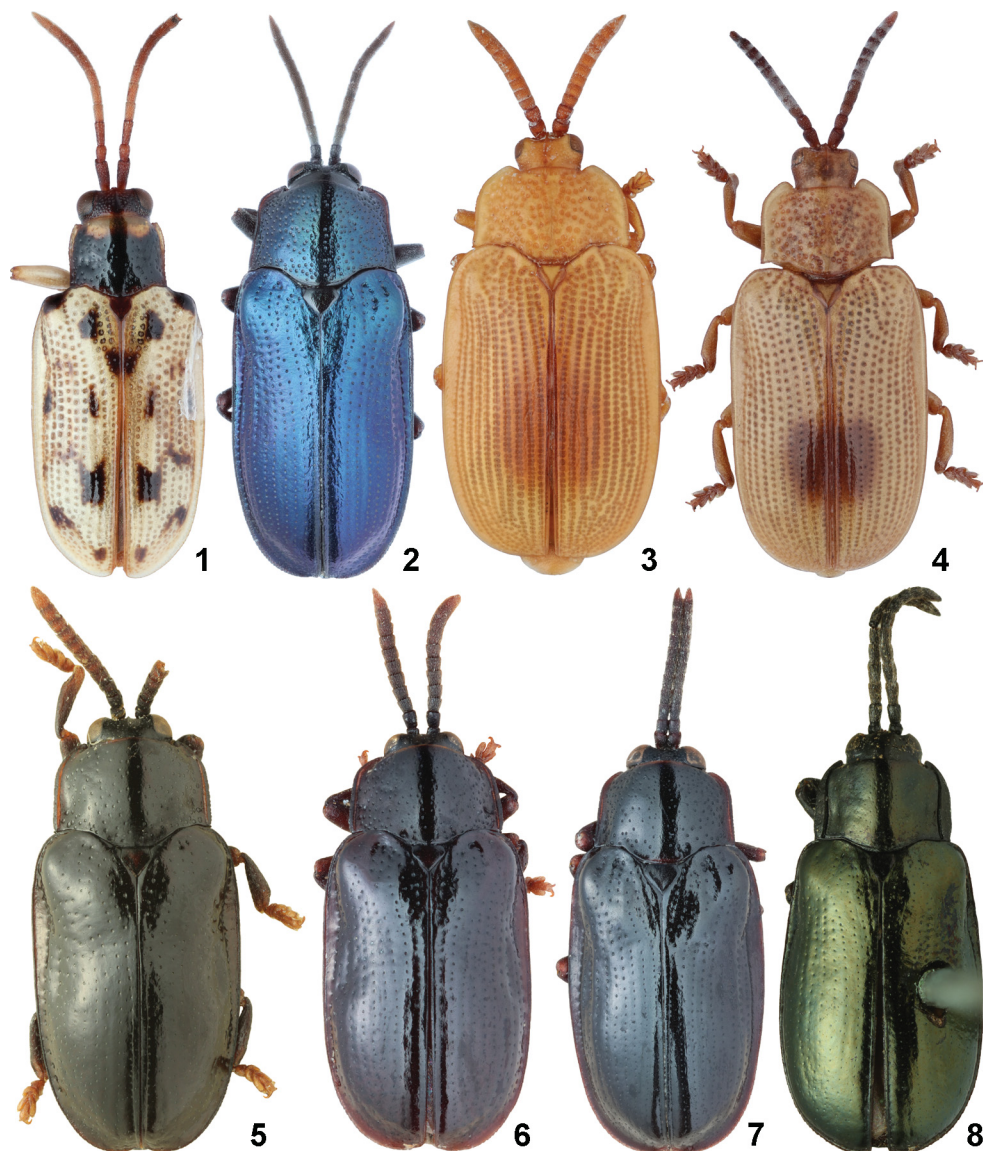
Serratispa quadricostata Staines, 2002 assign. nov.

(Fig. 12)

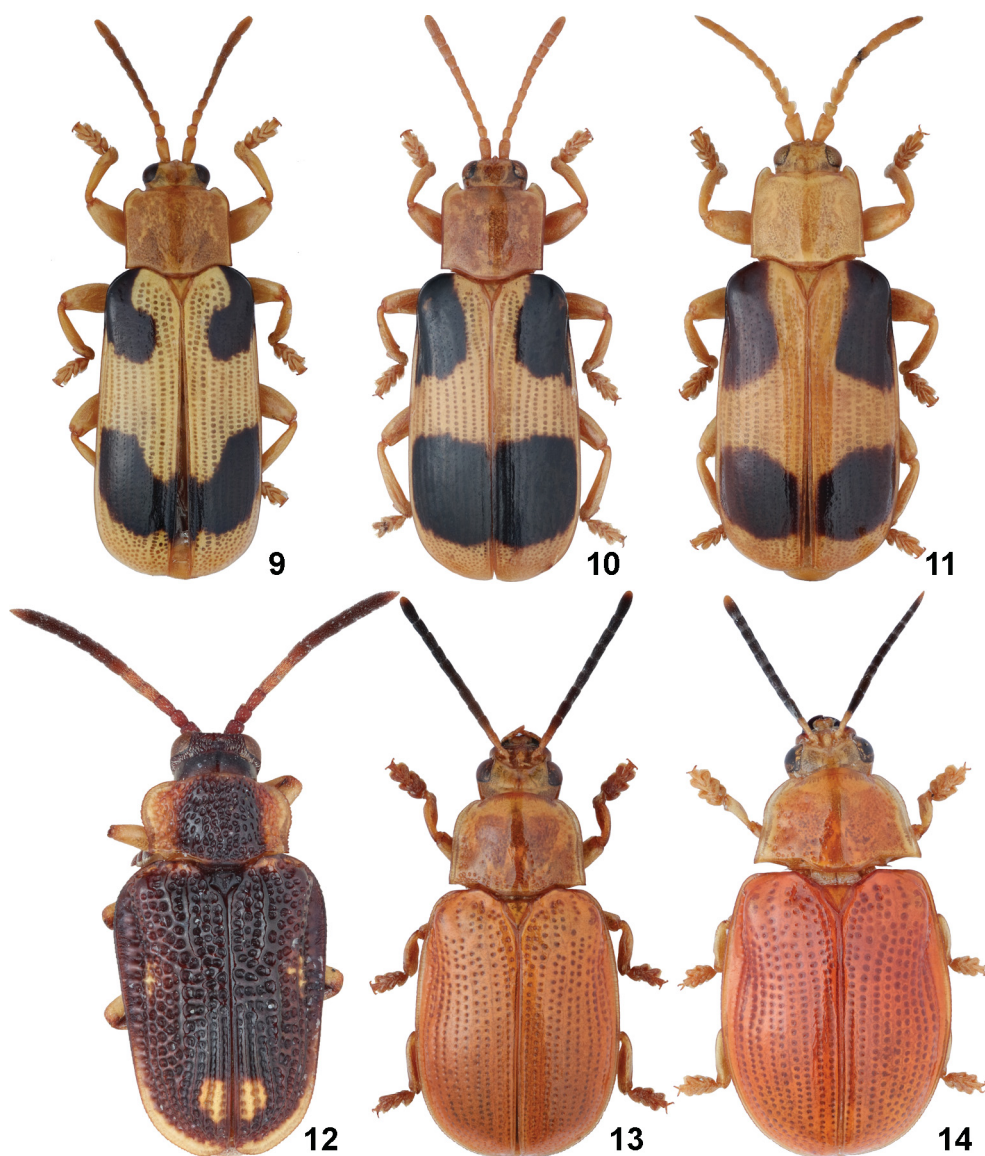
Serratispa quadricostata Staines, 2002: 749 (original description).

Type locality. Brazil, Amazonas, S o Paulo de Oliven a.

Type material examined. HOLOTYPE: glued, ‘S. o Paulo d’Oliven a | M. de Mathan | Juin.Juillet 1883. [w, p, cb, bf] || F. Monros | Collection | 1959 [w, p, cb] || HOLOTYPE | Serratispa [hw] | quadricostata [hw] | des. C. L. Staines 2001 [hw] [r, p, cb, bf] || BLNO | 002699 [blue, p, cb, bf] || USNM ENT | [data matrix code] | 00871554 [w, p, cb]’ (USNM).



Figs. 1–8. 1 – *Cephaloleia apertura* (Staines, 2013) (holotype, 4.3 mm); 2 – *Cephaloleia atripes* (Pic, 1926) (holotype, 5.1 mm); 3 – *Cephaloleia fernandoi* (Bondar, 1940) (paratype, LSPC, 6.3 mm); 4 – *Cephaloleia opaca* Baly, 1859 (Rio de Janeiro: Trinidad, LSPC, 6.0 mm); 5 – *Cephaloleia impressa* Uhmann, 1930 (holotype, 3.5 mm); 6 – *Demothispa clermonti* Pic, 1934 (holotype, 3.5 mm); 7 – *Cephaloleia jataiensis* (Pic, 1923) (holotype, 4.2 mm); 14 – *Xenispa aeneipennis* (Baly, 1859) (holotype, 3.7 mm).



Figs 9–14. 9 – *Cephaloleia fouquei* sp. nov. (holotype, 7.1 mm); 10 – *Cephaloleia renei* sp. nov. (holotype, 8.1 mm); 11 – *Cephaloleia kolbei* Weise, 1910 (Colombia: Río Dagua, LSPC, 9.5 mm); 12 – *Serratispa quadricostata* Staines, 2002 (holotype, 4.2 mm); 13 – *Xanthispa miniacea* (Blanchard, 1843) (Boivia: Potrerillo del Guenda, LSPC, 6.4 mm); 14 – *Xanthispa cimicoides* (Guérin-Méneville, 1844) (French Guyana: Le Larivot, LSPC, 6.5 mm).

Remarks. *Serratispa quadricostata* is the only species of this genus and both taxa were described by STAINES (2002) based on a single specimen from the Upper Amazon Basin. Staines placed the genus to the tribe Sceloenoplini without any justification. He stated only that ‘*Serratispa* differs from all other hispine genera by the following combination of characters: Antenna 11-segmented, filiform; head with vertex depressed, coarsely punctate; pronotum with a pair of setae on anterior margin behind head, lateral margins serrate; elytra costate with lateral plicae, with 11 rows of punctures at base.’ These characters unfortunately are not informative for tribal placement of *Serratispa*.

I have examined the holotype and in my opinion it does not belong to Sceloenoplini. The latter tribe is superficially similar to Chalepini in elytra with costae as remaining Neotropical tribes do not have primarily costate elytra. The costate elytra were probably one of the reasons why *Serratispa* was placed in Sceloenoplini, the other might have been presence of filiform antennae, however, Sceloenoplini have completely different formation of antennae. They always have terminal four antennomeres (VIII–XI) differently shaped or structured from the remaining antennomeres. *Serratispa* has the antennomeres VIII–XI similar to the four previous ones. Four other Neotropical tribes have pronotum with a seta present only near or at anterior corners: Arescini, Chalepini, Hispoleptini, and Imatidiini. Arescini clearly differs in transverse interantennal projection, large size, and enlarged middle coxae. Chalepini as well as Sceloenoplini differ in antennal insertions widely separated from mouthparts by a large clypeus. Hispoleptini have linear clypeus like *Serratispa* but differs in antennae inserted close to the vertex thus their base and entire antennomeres I are visible from above. In my opinion *Serratispa* belongs to Imatidiini as it has typical structure of the head for this tribe: vertex with longitudinal interantennal carina; antennae inserted close to the oral cavity thus their insertions and basal part of antennomere I are not visible from above, filiform, 11-segmented with only two basal sparsely pubescent antennomeres, remaining densely pubescent. Therefore I hereby transfer *Serratispa* to Imatidiini.

Serratispa runs to the couplet 9 in my key to genera of Imatidiini (SEKERKA 2014: 264), which includes *Pseudimatidium* and curious stout species currently classified in *Cephaloleia*. These clearly differ in smooth and convex vertex without carina and finely punctate elytra with entire margins while *Serratispa* has vertex deeply impressed on each side and divided by mid smooth carina and elytra very coarsely punctate with serrate margins.

Distribution. Brazil: Amazonas (STAINES 2002).

Xanthispa miniacea (Blanchard, 1843) comb. nov.

(Fig. 13)

Cephaloleia miniacea Blanchard, 1843: pl. 24, Fig. 1 (original illustration). BLANCHARD (1847): 211 (description).

Type locality. ‘province de Chiquitos (Bolivia)’.

Type material. Not studied, see Remarks.

Additional material examined. **BOLIVIA:** BENI: José Ballivián Province: Rurrenabaque, i.–ii.2000, 1 spec., C. Tello lgt. (MCSNM). **SANTA CRUZ:** Andrés Ibáñez Province: Reserva privada Potrerillos del Guendá, 17°40.26’S, 63°27.45’W, 370 m, 18.–25.xi.2011, 15 spec., D. Windsor & L. Sekerka lgt. (LSPC, MNKM, NMPC). **Ñuflo de Chávez Province:** Concepción-FCBC Alta Vista, 16°08.1’S, 61°56.1’W, 425 m, 28.xi.–5.xii.2011, 1 spec., D. Windsor & L. Sekerka lgt. (LSPC).

Remarks. WEISE (1910) transferred *C. miniacea* to the genus *Homalisp*a based on the original figure and description, and stated that it seems to be most similar to *Cephaloleia cimicoides* Guérin-Méneville, 1844. Baly (1859) did not mention *Cephaloleia miniacea* but proposed the genus *Xanthisp*a Baly, 1859 for *C. cimicoides* and differentiated it from *Homalisp*a by the shape of labium and relative lengths of the three basal antennomeres. WEISE (1910: 67–68) in a footnote considered *Xanthisp*a a subgenus of *Homalisp*a and subsequent authors accepted this classification (e.g. WEISE 1911, UHMANN 1957a, STAINES 2002). Most recently I have restored the genus status of *Xanthisp*a based on morphological characters (SEKERKA 2014).

Cephaloleia miniacea was validated by the figure 1 on plate 24 published on 15th December 1843 and all names are attributed to Emile Blanchard (BLANCHARD 1843, SHERBORN & GRIFFIN 1934). A short description was published four years latter by the same author but unfortunately its utility is limited to the colouration of the species: ‘Aurantiaco-miniacea, infra dilutior; antennis nigris, articulo primo rufo; pedibus rufo-miniaceis, tarsis piceis; elytris immaculatis, striato-punctatis. Long. 7 mill.’ [= Orange-like, underside paler; antennae black, antennomere I red; legs red-like, tarsi pitch black; elytra immaculate, striato-punctate. Length 7 mm.] (BLANCHARD 1847: 211).

The material from the *Voyage dans l’Amérique méridionale* by d’Orbigny was deposited in the MNHN. Some specimens were incorporated in the main collection (nowadays in boxes labelled as ‘collection historique’) while many others remained in the original boxes and are provided only with rounded green labels with number referring to the register of the Orbigny collection from his voyage to South America. In November 2016 I have searched all respective boxes and unfortunately did not find any specimen of *C. miniacea*. Nevertheless there is still possibility that the specimen was misplaced and exists somewhere in MNHN.

During recent research in Bolivia we found a species of *Xanthisp*a feeding on palms of the genus *Attalea* (Windsor & Sekerka, unpubl. data), which has similar colouration, size and shape of body as the original figure of *C. miniacea*. The new material was also collected from the same region as the type of the latter species. Therefore I assume our specimens belong to *C. miniacea* and hereby transfer it to the genus *Xanthisp*a.

Xanthisp miniacea and *X. cimicoides* are superficially quite similar but can be easily separated by the shape of pronotum, which is strongly trapezoidal in *X. cimicoides* (Fig. 14) and subquadratic and more or less parallel-sided at least in basal 2/3 in *X. miniacea* (Fig. 13). Also, *X. cimicoides* has broadly oval elytra in outline due to broader explanate margin of elytra while *X. miniacea* has narrower explanate margin of elytra and thus its body appears more ‘rectangular’.

Distribution. Bolivia: Beni: Santa Cruz (BLANCHARD 1847, present paper).

Xenisp aeneipennis (Baly, 1859) comb. nov.

(Fig. 8)

Cephaloleia aeneipennis Baly, 1859: 59 (original description).

Type locality. ‘Venezuela’.

Type material examined. HOLOTYPE: pinned, ‘Type | H. T. [w, p, cb, circle label with red frame] || Type [g, hw, s] || Cephaloleia | aeneipennis | Baly | Venezuela [w, hw by Baly, s]’ (BMNH).

Remarks. BALY (1859) described this species based on a single specimen as he clearly indicated. Subsequently it was mentioned only in catalogues (e.g. WEISE 1911, BRYANT 1942, UHMANN 1957a). STAINES & GARCÍA-ROBLEDO (2014) provided redescription, colour photograph and reported additional specimen from Colombia without any further data.

I have studied the holotype and in my opinion the species belongs to the genus *Xenispa* as it possesses all principal characters of that genus, e.g. long antennae, serrate lateroapical margins of elytra, smooth and convex frons and vertex. Therefore I hereby transfer *Cephaloleia aeneipennis* to the genus *Xenispa*.

Xenispa aeneipennis is most similar to *X. carinata* (Pic, 1934) also known from Venezuela. The two species can be distinguished using the herein proposed key to Venezuelan species of *Xenispa*, see page 374.

I was not able to verify the identity of the specimen from Colombia (STAINES & GARCÍA-ROBLEDO 2014) as it was not found in the collections of NHMW (Harald Schillhammer, pers. comm. October 2017) and I doubt that the identification was correct since the authors made numerous errors in species identification. So the holotype remains the only specimen of this species known to me.

Distribution. Colombia? (STAINES & GARCÍA-ROBLEDO 2014) and Venezuela (BALY 1859).

Xenispa argentina (Monrós & Viana, 1947)

(Fig. 19)

Demothispa argentina Monrós & Viana, 1947: 158 (original description).

Type locality. 'Argentina: Formosa, dto Capital: Isla de Oro'.

Type material examined. HOLOTYPE: glued (missing right midtarsi and left hind leg), 'Formaso | M. Viana | I. 936 [w, hw, cb] || 55 [red, typed on typing machine, cb] || Dibujado [w, hw, cb] || Demothispa [Uhmman hw] | sp. [Uhmman hw] | UHMANN DET.37 [w, p, s] || ARGENTINA | 1968 Colin. | J. Daguerre [w, p, cb] || Demothispa [Vianna hw] | argentina nov. [Vianna hw] | M. VIANA DETRM. [w, p, cb] || 9 [g, typed, cb] || Holotipo [r, hw, cb] || HOLOTYPE [r, p, cb, bb] || BLNO | 002832 [blue, p, cb, bb] || USNMMENT | [data matrix barcode] | 00911217 [w, p, cb]' (USNM).

Remarks. MONRÓS & VIANA (1947) described *D. argentina* based on two specimens and compared it with *D. latifrons* and *D. bicoloricornis*. STAINES (2009) transferred it to the genus *Stilpnaspis* Weise, 1905 without any explanation. Most recently, I have assigned it to *Xenispa* Baly, 1859 based on original figure but stated that the transfer is tentative as no type material was examined. Subsequently, I had opportunity to study the holotype of *D. argentina* (Fig. 19) and I can hereby confirm the transfer to *Xenispa* as it has all the principal characters of that genus as mentioned in my previous paper (SEKERKA 2014).

Xenispa argentina has similar shape of body to many other species of that genus but differs at first glance by uniformly yellow dorsum while all other species of *Xenispa* have at least elytra black or metallic blue. The species is so far known only from the two type specimens.

Distribution. Argentina: Formosa and Paraguay: Itapúa (MONRÓS & VIANA 1947).

Xenispa atra (Pic, 1926a)

(Fig. 15)

Demothispa atra Pic, 1926a: 13 (original description).

Cephaloleia cyanea Staines, 1996: 25 (original description), **syn. nov.**

Cephaloleia coroicoana (misidentification): STAINES & GARCÍA-ROBLEDO (2014): 117 (partim, faunistics).

Type localities. *Demothispa atra*: ‘Venezuela’. *Cephaloleia cyanea*: Venezuela, Aragua State, Rancho Grande, 1100 m.

Type material examined. *Demothispa atra*: HOLOTYPE: glued, ‘Venezuela [w, hw by Pic, s] || 9 [w, hw, s] || Gestro vidit [w, hw by Pic, s] || coll Pic [w, hw by Pic, s] || voi | aeneipennis | Baly [w, hw by Pic, s] || type [y, hw by Pic, s] || TYPE [r, p, cb] || Museum Paris | Coll. M. Pic [b, p, cb] || C. atra | n sp [w, hw by Pic, s]’ (MNHN).

Cephaloleia cyanea: HOLOTYPE: glued, ‘VENEZUELA: Aragua | Rancho Grande | 8 [hw] May 1973 1100 m. | Ginter Ekis [w, p, cb] || Type Number | 105427 [hw] | U.S.N.M. [r, p, cb] || HOLOTYPE | Cephaloleia [hw] | cyanea Staines [hw] | Des. C. L. Staines [hw] || 1994 [hw] [r, p, cb] || USNM ENT | [data matrix code] | 00871337 [w, p, cb]’ (USNM).

Additional material examined. **BRAZIL**: ‘Brasilia’ without further data, 1 spec. (BMNH, ex coll. A. Fry, register no. 43830). **VENEZUELA: ARAGUA**: Rancho Grande Biological Station, 10°21’N, 67°41’W, 1300 m, 12.–14.v.1998, 1 spec. (Malaise trap), J. Ashe, R. Brooks & R. Hanley lgt. (SEMC).

Remarks. Pic (1926a) described *D. atra* presumably based on a single specimen as the description is written in singular and only a single length measurement was mentioned, and compared it to *D. jataiensis*. I moved *D. atra* tentatively to *Xenispa* based on the primary description (SEKERKA 2014) as the holotype was on loan. In November 2016 I had opportunity to study the holotype and those observations indicate the species clearly belongs to *Xenispa* as it possesses all principal characters of that genus, e.g. long antennae, serrate apical margin of elytra, smooth and convex frons and vertex.

STAINES (1996) described *Cephaloleia cyanea* based on series of eleven specimens: holotype was collected in Rancho Grande in Venezuela, three paratypes in Caracas (USNM), one paratype near Arcabuco in Bocaya Department of Colombia (USNM), and remaining six paratypes in Monte Verde in Costa Rica (CMNC and collection of Henry Hespenheide, USA). I had opportunity to study most of the type series and found none of the paratypes is conspecific with the holotype and each belongs to one of three different species. The original description is a mixture of characters of all four taxa but seems to be mostly based on specimens from Caracas. One of them was also figured in the original description (STAINES 1996). I have compared the holotype of *C. cyanea* with that of *D. atra* and found that they are conspecific and thus I synonymize *C. cyanea* with the latter species.

STAINES & GARCÍA-ROBLEDO (2014) published a redescription of *C. cyanea* but it was mostly based on the Colombian specimen, which they also figured. The material listed in ‘Specimens examined’ consists of actual paratypes designated in 1996. STAINES & GARCÍA-ROBLEDO (2014: 117) also recorded *Cephaloleia coroicoana* Uhmann, 1930 from Venezuela based on a single specimen. However, the specimen was misidentified and belongs to *X. atra* and is given here in additional material examined. Therefore, Venezuela must be removed from the range of *C. coroicoana*. *Xenispa atra* is known to me based on four specimens only, the two types and two additional specimens listed here. The BMNH specimen has vague locality data ‘Brasilia’ and came from the collection of W. W. Saunders. I think it was quite likely mislabelled and the species does not occur in Brazil as the specimen is morphologically identical to the remaining specimens not displaying any morphological variability. According to the properly labelled specimens the species occurs in the central part of Cordillera de la Costa in Venezuela, which has a quite unique flora and fauna with high local endemism. Therefore I do not include Brazil in the species distribution.

Xenispa atra is a very characteristic species as it has narrow lateral margins of pronotum while most *Xenispa* species have wide lateral margins. This species can be easily separated from other Venezuelan species of *Xenispa* using the key on page 374.

Distribution. Venezuela: Aragua (STAINES 1996).

Xenispa fouquei sp. nov.

(Fig. 18)

Type locality. Venezuela, State of Mérida, Mérida-La Estación La Montaña, approx. 8°34.433'N, 71°6.879'W, 2500 m a.s.l.

Material examined. HOLOTYPE: ♂, glued, 'VENEZUELA Es[t]ado Mérida | 2500m above La Montaña | S. above Mérida | on bamboo 6 Aug 1977 | Adams/Bernard [w, p, cb] || Adams/Bernard | BMNH{E} | 1978-281 [w, p, cb]' (BMNH). PARATYPES: **VENEZUELA:** 3 ♂♂ 6 ♀♀, same data as holotype (BMNH, 1 ♂ 1 ♀ LSPC, 1 ♂ NMPC). 2 spec., 'VENEZUELA | Merida, Carbonera | 2000m 19.iv.1989 [w, p, sl]' (DSCC, LSPC); 1 spec., 'Venezuela, Mérida | Carbonera, 2300 m | 17.VI.1993, leg. C. Bordón [w, p, cb]' (MDVC); 2 spec., 'Venezuela – Tachira | La Grita, 2400 m | 16.III.1983, leg. Bordon [w, p, cb]' (LSPC); 2 spec., 'PREGONERO, m. | 2500. Edo.Táchira [w, p, cb] || VENEZ. Bordón | leg. 2 XI 1978 [w, p, cb]' (MDVC); 1 spec., 'carret. Trujillo- | Boconé, 2500 m. [w, p, cb] || Edo.Trujillo.VENEZ | Bordón 17 XI 1981 [w, p, cb]' (LSPC).

Description. Body broadly oval. Length: 5.8–7.2 mm.

Head, pronotum, pro- and mesothorax yellow. Scutellum, metathorax and abdomen black. Antennomere I brown, remaining black. Fore and middle legs yellow including trochanters and coxae and infusate on knees. Hind legs infusate brown-black, tarsi, trochanters and coxae pale brown. Elytra dark metallic blue.

Frons and vertex smooth, shiny, and impunctate. Vertex with moderately deep oval basal impression. Eyes large, gena clearly visible, smooth. Interantennal carina present, short and low. Mouthparts directing forwards but not visible from above. Antennae long, thin, with two basal sparsely pubescent antennomeres, remainder densely pubescent. Length ratio of antennomeres: 100 : 103 : 208 : 164 : 155 : 145 : 139 : 143 : 132 : 117 : 200. Antennomere III as long as I and II combined, from IV antennomeres gradually shorter toward apex of antenna except XI, which is twice as long as I. Antennomere I globose, remaining antennomeres elongate, distinctly longer than broad. Tip of antennomere XI subacuminate. Antennomeres unmodified.

Pronotum sub-trapezoidal, 1.75× as wide as long (measured at base), with rounded sides. Basal margin bisinuate, basal corners acute. Anterior margin deeply emarginate above head. Anterior corners projecting anterad, wide and broadly rounded. Anterior margin gradually bent towards corners forming obtuse angle. Tubercles with setigerous pores situated in the angle. Lateral margins moderately explanate, not distinctly bordered from disc. Outer edge moderately swollen, moderately canaliculate and slightly serrate in basal 1/2. Disc smooth, finely and very sparsely micro-punctate. Lateral slopes regular, not impressed, each with circa 15 coarse and sparsely arranged punctures.

Scutellum subpentagonal, smooth, shiny, and microsculptured.

Elytra drop-shaped, widening apically. Base wider than base of pronotum, bisinuate, smooth, humeral angles broadly rounded. Humeral calli convex and impunctate. Punctuation regularly arranged in ten rows. Scutellar row present, long, composed of circa 15 punctures and stretching from base to postscutellar hump. Overall, punctuation of elytra moderately coarse. Punctures in rows densely and more or less regularly arranged. Inner rows less impressed, more densely punctate with smaller and less regular punctures, interspaces mostly narrower than puncture diameter. Outer rows gradually more impressed towards explanate margin, punctures larger and more sparsely arranged, interspaces 1–2× as wide as puncture diameter. Ultimate and penultimate rows of punctures visible in whole length and without distinct vacancies. Intervals circa 5–6× as wide as puncture diameter, inner ones slightly narrower. All intervals micro-reticulate and sparsely micropunctate (at magnification 90×).

Intervals appear more or less elevated due to impressed rows of punctures. Explanate margin broad, approximately as wide as 1/4 of disc, appears slightly rugose due to irregular surface, shiny and impunctate. Outer margin moderately swollen, distinctly canaliculate and dentate from humeral angles to apex. Each denticle with short seta on tip. Apices conjointly rounded, slightly emarginate in females.

Prosternal collar smooth and shiny. Prosternal process broad, strongly expanded apically and impressed along whole length. Epipleura smooth, micro-reticulate and sparsely pubescent. Thoracic ventrites micro-reticulate, sparsely punctate and pubescent. Mesepisterna micro-reticulate, densely punctate and pubescent. Abdominal ventrites micro-reticulate, moderately densely punctate and pubescent with long semiadherent setae. Ventrites I and II centrally fused.

Sexual dimorphism indistinct. Ventrite V emarginate in males and truncate in females.

Legs normal, last tarsomere elongate, projecting beyond sole of penultimate. Claws simple, divergent.

Comparative diagnosis. The new species is morphologically similar only to *X. pulchella* Baly, 1859, the type species of the genus. They both share similar shape of pronotum and body but can be distinguished by characters summarized in the Table 2. Venezuelan species of *Xenispa* can be distinguished using key on page 374.

Etymology. The species is dedicated in loving memory to René Fouqué (1980–2016), friend and enthusiastic entomologist, who was world specialist on the tenebrionid tribe Stenosini.

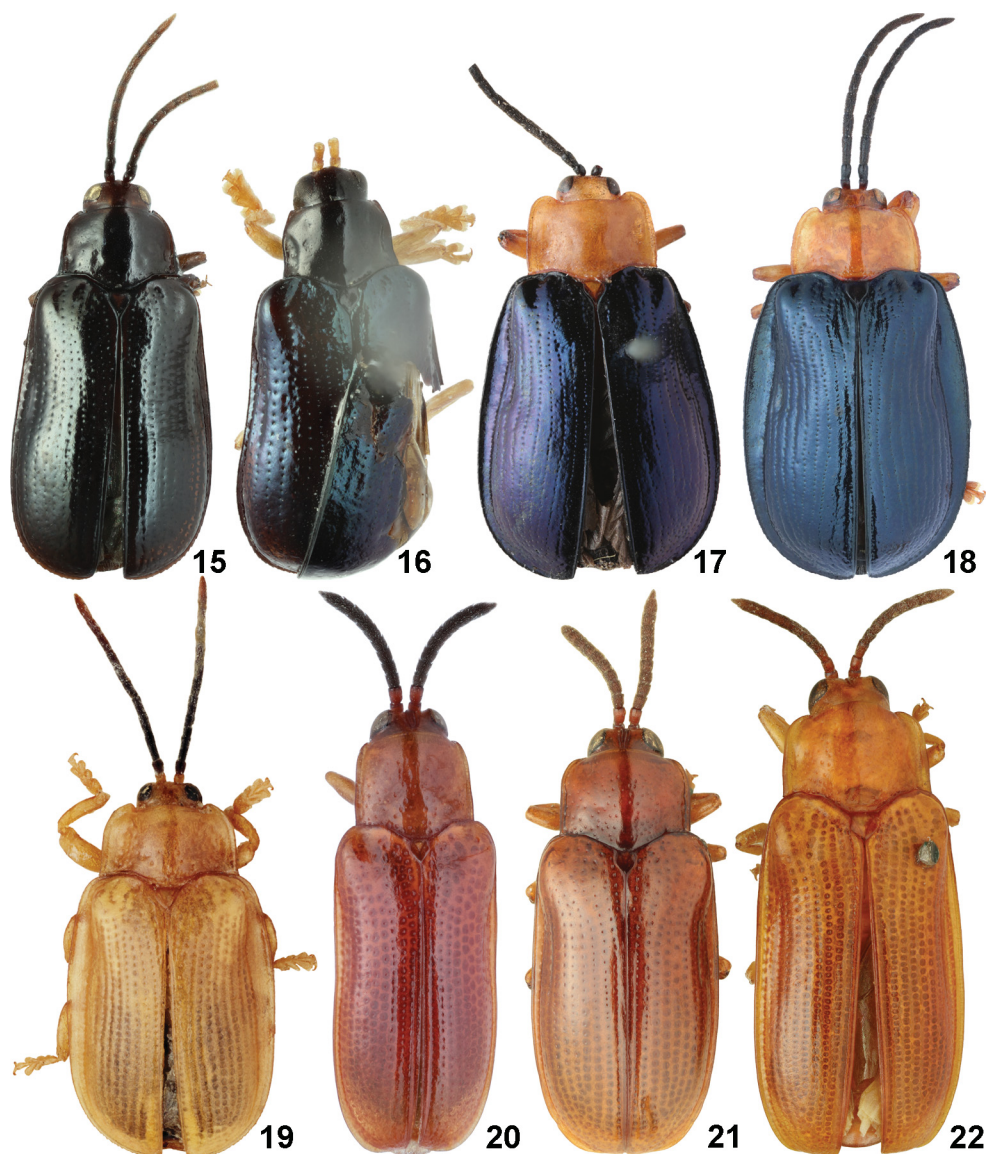
Biology. According to the label data the BMNH specimens were collected on bamboo; what is most likely correct as species of *Xenispa* are mainly associated with various Neotropical bambusoid Poaceae.

Remarks. According to the label data the BMNH specimens were collected about fifty meters in elevation above La Montaña, which is the middle station of the cable tram connecting Barinitas and La Aguada. The coordinates are approximate and do not represent the actual spot where the specimens were collected. They were taken using GoogleEarth on the cable tram transect at 2500 m a.s.l. just to illustrate habitat.

Distribution. Venezuela: Mérida.

Table 2. Distinguishing characters between *X. pulchella* Baly, 1859 and *X. fouquei* sp. nov.

character / species	<i>Xenispa pulchella</i> Fig. 17	<i>Xenispa fouquei</i> sp. nov. Fig. 18
pronotum shape	sub-rectangular and parallel-sided	narrowing anteriorly
anterior corners of pronotum	broad and rounded	narrow and subangulate
pronotum: lateral sides of disc	deeply impressed	not impressed
explanate margin of elytra	narrow	broad
punctuation of elytra	rows on the same level as intervals, punctures smaller	rows distinctly impressed, punctures larger, overall punctuation coarser
scutellum	yellow	brownish-black
tarsi	black	yellow



Figs 15–22. 15 – *Cephaloleia cyanea* Staines, 1996 (holotype, 4.0 mm); 16 – *Xenispa gilvipes* (Uhmann, 1930) (holotype, 4.4 mm); 17 – *Xenispa pulchella* Baly, 1859 (holotype, 7.7 mm); 18 – *Xenispa fouquei* sp. nov. (♂ paratype from the type locality, 6.9 mm); 19 – *Xenispa argentina* (Monrós & Viana, 1947) (holotype, 5.1 mm); 20 – *Hybosipa delectabilis* (Staines, 1996) (holotype, 3.9 mm); 21 – *Hybosipa sulciceps* (Baly, 1885) (syntype, BMNH, 3.1 mm); 22 – *Hybosipa truncatipennis* (Baly, 1869) (holotype, 6.75 mm).

***Xenispia gilvipes* (Uhmman, 1930) comb. nov.**

(Fig. 18)

Cephalolia gilvipes Uhmman, 1930b: 230 (original description).**Type locality.** Costa Rica, San José Province, Santa María de Dota, approx. 9°39'N, 83°58'W, ca. 1500–1600 m.**Type material examined.** HOLOTYPE: pinned (both antennae missing, right elytron partly broken), 'COSTA RICA | F. NEVERMANN | 4.I.24 [g, p, cb, last row hw] || Sta. Maria de | Dota | an Blüten [verso of the previous label, hw] || Type No. | 54603 [hw] | USNM [r, p, cb] || HOLOTYPE [r, t, s] || Cephalolia | gilvipes n. sp. [w, hw by Uhmman, s] || USNM ENT | [data matrix code] | 00871395 [w, p, cb]' (USNM).**Additional material examined.** COSTA RICA: CARTAGO: Cañon, 2450 m, 28.xi.1994, 22 spec., H. Forster lgt. (15 spec. NHMW, 6 spec. LSPC, 1 spec. NMPC).

Remarks. UHMANN (1930b) described this species based on a single specimen and stated that it was collected on flowers. The holotype is strongly damaged missing both antennae from the antennomere II on and right elytron is broken due to the pin. In my opinion this species does not belong to *Cephaloleia* as it has conspicuously serrate apical margin of elytra, long antennae compared to the length of body and smooth convex frons and vertex. These characters are typical for the genus *Xenispia* and *C. gilvipes* conforms to other species currently classified in that genus. Therefore I hereby transfer *C. gilvipes* to *Xenispia*.

The species was so far reported only three times based on specimens collected in high mountains of central Costa Rica above 1500 m a.s.l. (FLOWERS & HANSON 2003, McKENNA & FARELL 2005, STAINES & GARCÍA-ROBLEDO 2014). I had no opportunity to verify any of the recently published specimens, thus I cannot confirm their identifications. The specimens reported here agree perfectly with the holotype, having the same shape, colouration and punctuation. Particularly characteristic is the shape of pronotum as it has only very narrow lateral margins, which are not explanate. A similarly shaped pronotum is known only in *Xenispia uhmanni* (Pic, 1934) from Colombia, which differs in having black antennae and legs while *X. gilvipes* has them pale brown. The locality Cañon is situated on the Pan-American Highway and is about 6 km air distance to NW from the type locality of *X. gilvipes*.

Distribution. Costa Rica: Cartago, Limón, Puntarenas (STAINES & GARCÍA-ROBLEDO 2014, present paper), Heredia (McKENNA & FARELL 2005), and San José (FLOWERS & HANSON 2003).

Key to Venezuelan species of *Xenispia*

- 1 Pronotum of the same colour as elytra, legs black or metallic. Small species, length below 4 mm with narrow margin of elytra. 2
- Pronotum, head, fore- and mid-legs yellow. Large species, length 6.0–7.5 mm with broadly explanate margin of elytra (Fig. 18). ***X. fouquei* sp. nov.**
- 2 Lateral margins of pronotum strongly swollen, broad and separated by a deep sulcus from the disc. Dorsum metallically coloured. 3
- Lateral margins of pronotum narrow and separated by shallow sulcus from the disc. Dorsum pitch-black, pronotum somewhat paler coloured (Fig. 15). ***X. atra* (Pic, 1926)**
- 3 Pronotum narrower, 1.4–1.5× as wide as long. Swollen lateral margin of pronotum of the same width, narrowing only anteriorly. Elytra black with greenish or antracite metallic tint. 4

- Pronotum broader, 1.7× as wide as long. Swollen lateral margin of pronotum distinctly narrowed at base and anteriorly. Elytra very dark brown with bluish metallic tint. *Xenispa* sp.¹
- 4 Pronotum shiny, microreticulation centrally vanished and more or less visible only laterally. Elytra metallic antracite. *X. carinata* (Pic, 1934)
- Pronotum subopaque due to more prominent microreticulation. Elytra very dark olive-green (Fig. 8). *X. aeneipennis* (Baly, 1859)

Hybosispini Weise, 1910

WEISE (1910) proposed the tribe for a single species *Hybosispa melanura* Weise, 1910 from Bolivia which is externally very similar to *Cephaloleia* but has no seta on pronotum while all Imatidiini have a seta at each anterior corner of pronotum or next to them. *Hybosispa* also has strongly convex body, which is nearly circular in cross-section, and head with prominent carina on inner margin of the eye. Imatidiini have more or less flattened body, always with transverse cross-section and head without prominent carina along eyes.

Latter on, UHMANN (1933, 1939, 1940) described three more *Hybosispa* species from Brazil. Subsequently, UHMANN (1964: 405) and STAINES & GARCÍA-ROBLEDO (2014: 7) transferred *Cephaloleia macella* Pic, 1923 and *C. bipartita* Pic, 1926, respectively, to *Hybosispa*. Finally, SEKERKA (2014) revised genera of Imatidiini and found that *Solenispa* Weise, 1905 lack setae on pronotum and have similar morphology of the head to *Hybosispa* and thus transferred *Solenispa* to Hybosispini. WEISE (1905) when describing *Solenispa* mentioned the shape of the head and based on it differentiated *Solenispa* from *Stenispa* Baly, 1859, however, he did not notice that *Solenispa* lacks pronotal setae.

I have studied type material of various species of *Cephaloleia* and found three more species lack pronotal setae and have the shape of body and formation of the head similar to *Hybosispa* and are here transferred to the latter genus. Additionally, the only non-Andean member of the genus *Solenispa* – *S. claripes* Pic, 1923 is here transferred also to *Hybosispa*. Therefore *Hybosispa* presently contains ten species and they can be recognized using a key proposed on page 377.

Hybosispa claripes (Pic, 1923) comb. nov.

Solenispa claripes Pic, 1923: 10 (original description).

Type locality. Original type locality ‘Brésil’ specified by DESCARPENTRIES & VILLIERS (1959) to Brazil, Rio de Janeiro, Theresópolis based on the label data of the type.

Type material examined. HOLOTYPE: glued, ‘Therezopolis [g, hw by Donckier, cb] || *Solenispa* | *claripes* m [w, hw by Weise, s] || co-Type [w, p in red, cb] || type [y, hw by Pic, s] || TYPE [r, p, cb] || *Sol. claripes* | (1923) Pic [w, hw by Pic, s]’ (MNHN).

Remarks. According to the label data the specimen was studied by J. Weise who recognized it as a new species of *Solenispa*, however, he did not publish the description and PIC (1923)

¹ This refers to the three paratypes of *X. cyanea* Staines, 1996 from Caracas, one of which was figured by STAINES (1996). They possibly belong to a new species, however, all three have broken and missing antennae therefore I rather avoid describing this taxon until I have better preserved specimens.

validated the name by very short and uninformative description, which is written in singular and Pic gave only a single length measurement suggesting that he had only a single specimen at his disposal. Afterward it was listed only in catalogues (e.g. UHMANN 1957a, 1964).

I have studied the holotype and in my opinion it belongs to the genus *Hybosispa* as it has typical shape of that genus and abdominal ventrites are not pubescent, while all known species of *Solenispa* have them densely pubescent. Therefore I hereby transfer *S. claripes* to *Hybosispa*.

It is most similar to *H. nitida* Uhmman, 1939 and *H. rufiventris* Uhmman, 1940 as all three share uniformly black dorsum. However, *H. claripes* differs in more impressed frons and vertex and pale coloured legs.

Distribution. Brazil: Rio de Janeiro (DESCARPENTRIES & VILLIERS 1959).

***Hybosispa delectabilis* (Staines, 1996) comb. nov.**

(Fig. 20)

Cephaloleia delectabilis Staines, 1996: 26 (original description incl. black-and-white photograph).

Type locality. Mexico, Chiapas, Parque educativo Laguna Belgica, 16 km NW of Ocozacoatlán.

Type material examined. HOLOTYPE: glued, 'MEXICO, Chiapas, | Pq. Laguna Belgica, | 16kmNW Ocozacoatlán | 14.VI.1989.H.Howden [w, p, cb] || Flight intercept | trap [w, p, cb] || HOLOTYPE | Cephaloleia [hw] | delectabilis [hw] | Staines 1994 [hw] | Des. C. L. Staines [hw] [r, p, bc] || [data matrix code] | Canadian Museum of | Musée canadien de la | NATURE | CMNEN 00012091 [w, p, cb, bf]' (CMNC).

Remarks. STAINES (1996) described *C. delectabilis* based on single specimen and placed it in the genus *Cephaloleia* without further comment. Studying the black-and-white photograph, which accompanied the original description I doubted that the species belongs to the genus *Hybosispa* but essential characters were not visible or mentioned in the original description. I loaned the holotype and found that it has no seta on anterior margin of pronotum and the inner margins of eyes are produced, therefore I hereby transfer the *C. delectabilis* to *Hybosispa*. The species is so far known only from the holotype.

Distribution. Mexico: Chiapas (STAINES 1996).

***Hybosispa sulciceps* (Baly, 1885) comb. nov.**

(Fig. 21)

Cephaloleia sulciceps Baly, 1885: 26 (original description).

Type locality. Panama, Chiriquí Province, Bugaba.

Type material examined. SYNTYPE: glued, 'SYN- | TYPE [w, p, s, circular label with blue frame] || Bugaba, | Panama. | Champion. [w, p, cb] || Godman-Salvin | Coll., Biol. | Centr.-Amer. [w, p, cb]' (BMNH).

Remarks. BALY (1885) described *C. sulciceps* based on unknown number of specimens. I assume that he must have had longer series of specimens because he did not state precise number and he mentioned both sexes. In the *Biologia Centrali Americana* when Baly had only a limited number of specimens, he usually mentioned exactly how many he got for description while in the case of a long series he seldom did that. I have studied syntypes preserved in BMNH and all specimens lack setae on pronotum, therefore I hereby transfer *C. sulciceps* to *Hybosispa*. I have no doubt that all specimens belong to a single species, thus there is no need for lectotype designation. As type material examined is listed only as a single syntype which I had on loan, this one is figured here (Fig. 21).

The exact position of the type localities is unknown. Bugaba without information on elevation usually refers to lowland (below 300–500 ft) part of the Bugaba District in the Western Chiriquí. The area is situated around present day city of La Concepción and the small pueblo Bugaba is situated ca. 3.7 km to south. Some other syntypes in BMNH and a syntype in USNM have on the label also elevation 800–1500 ft. This elevation, however might refer to much larger area extending from South-western slopes of Volcán Barú to Río Chiriquí Viejo. Based on CHAMPION's (1907) itinerary he visited several places in this area but in the end specimens were provided just with label 'Bugaba' and elevation.

STAINES (1996) published additional 17 specimens of this species from Panamá and Costa Rica. These records were partly published again by STAINES & GARCÍA-ROBLEDO (2014) and mixed with several new records. Thus far I have had no opportunity to verify these records.

Distribution. Costa Rica: Cartago, Limón, Punarenas and Panamá: Chiriquí, Panamá (BALY 1885, STAINES 1996).

Hybosispa truncatipennis (Baly, 1869) comb. nov.

(Fig. 22)

Cephaloleia truncatipennis Baly, 1869: 371 (original description).

Type locality. 'Upper Amazons'.

Type material examined. HOLOTYPE: pinned, 'Type | H.T. [w, p, cb, circular label with red frame] || Baly Coll. [w, p, cb] || Cephaloleia | truncatipennis | Baly | Upper Amazons [g, hw by Baly, cb]' (BMNH).

Remarks. BALY (1869) described *C. truncatipennis* presumably based on a single specimen as he gave just a single length and mentioned only a female. The species was only listed in catalogues since its description until STAINES & GARCÍA-ROBLEDO (2014) who included it in their revision of *Cephaloleia* and reported one specimen from Capella in Brazil. However based on the photograph presented in the revision (STAINES & GARCÍA-ROBLEDO 2014: 314, Fig. 258) the specimen was misidentified and belonged to another species of *Cephaloleia*. The redescription of *C. truncatipennis* is also not in full agreement with the holotype and seems to combine characters of both specimens thus may not be reliable.

The holotype (Fig. 22) has anterior corners of pronotum clearly without any seta and head just like the other species of *Hybosispa* therefore I hereby transfer it to the latter genus. The species is thus far known only from the single specimen collected by H. W. Bates in the Upper Amazons. Unfortunately no precise data are available for it and thus whether it was captured in Brazil or in Peru is unknown.

Distribution. Amazonas state of Brazil or Region of Loreto in Peru.

Key to species of *Hybosispa*

- 1 Abdominal ventrites bare; antennae short and thick, distal antennomeres as wide as long. Lowland or mid-elevation species (usually below 1000 m) from southern Mexico to southern Brazil. *Hybosispa* Weise, 1910 ... 2
- Abdominal ventrites densely pubescent; antennae longer and slender, most of distal antennomeres longer than wide. Eight species in the Cordillera in Central America and

- the Andes from Costa Rica to Bolivia; all species at elevations above 1200 m. *Solenispa* Weise, 1905
- 2 Dorsum uniformly black. 3
- Dorsum at least partly red. 5
- 3 Legs and antennae dark coloured, pitchy-brown; frons weekly impressed with low and obtuse medial carina reaching only anterior part of vertex. 4
- Legs and antennomere I pale coloured, yellowish-brown; frons deeply impressed between eyes with sharp and high mid carina extending to whole width of vertex. Brazil: Rio de Janeiro. *H. claripes* (Pic, 1923)
- 4 Abdomen black; pronotum parallel-sided in whole length not converging in anterior third. Brazil: Santa Catarina. *H. nitida* Uhmann, 1939
- Abdomen reddish-brown; sides of pronotum rounded in anterior third and distinctly converging anteriad. Brazil: Santa Catarina. *H. rufiventris* Uhmann, 1940
- 5 At least apical part of elytra black. 6
- Dorsum uniformly red. 9
- 6 Head black; inner margin of eyes not strongly carinate and projecting anteriad. 7
- Head red; inner margin of eyes forming sharp carina, which is projecting anteriad. ... 8
- 7 Pronotum 1.6–1.7 times as wide as long with conspicuously flat antero-lateral sides. Brazil: Rio de Janeiro. *H. strandi* Uhmann, 1933
- Pronotum 1.3–1.4 times as wide as long, not flattened on sides. Bolivia: Cochabamba.. *H. macella* (Pic, 1923)
- 8 Pronotum subrectangular, for most length parallel-sided and tapering anterad only at anterior corners; abdomen and thorax mostly black. Peru: Cuzco. *H. bipartita* (Pic, 1926)
- Pronotum in anterior third rounded distinctly tapering anterad; ventral side red. Bolivia: La Paz and Peru: Marcapata. *H. melanura* Weise, 1910²
- 9 Smaller species, body length below 4 mm. Central-American species. 10
- Larger species, length 5.7 mm. Described from ‘Upper Amazons’. *H. truncatipennis* (Baly, 1869)
- 10 Pronotum transverse, 1.4 times as wide as long; distal antennomeres brown. Costa Rica and Panamá. *H. suliceps* (Baly, 1885)
- Pronotum subquadrate, 1.1 times as wide as long; distal antennomeres black. Mexico: Chiapas. *H. delectabilis* (Staines, 1996)

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² UHMANN (1957a) recorded *H. melanura* as new to Peru based on specimen(s) from Marcapata, which is the type locality of *H. bipartita*. I did not examine the material, thus I cannot verify it but it might be that the record from Marcapata belongs to *H. bipartita*. On the other hand the two taxa are quite similar and further studies based on more material are needed to verify whether they are two distinct species or just forms of a single species.

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References

- BALY J. S. 1859: *Catalogue of Hispidiae in the collection of the British Museum. Part I*. Trustees of the British Museum, London, 172 pp. + 9 pls.
- BALY J. S. 1869: Descriptions of new genera and species of Hispidiae; with notes on some previously described species. *Transactions of the Entomological Society of London* **1869**: 363–382.
- BALY J. S. 1885: Hispidiae. Pp. 1–72. In: GODMAN F. D. & SALVIN O. (eds): *Biologia Centrali-Americana. Insecta. Coleoptera. Vol. VI. Part 2. Phytophaga (part)*. Taylor & Francis, London, x + 249 pp. + 13 pls.
- BLANCHARD E. 1843: [*Cephaloleia miniacea*]. Pl. 24, Fig. 1. In: d'ORBIGNY A. 1837–45 (ed.): *Voyage dans l'Amérique méridionale (Le Brésil, la République Orientale de l'Uruguay, la République Argentine, la Patagonie, la République du Chili, la République de Bolivie, la République du Pérou), exécuté pendant les années 1826, 1827, 1828, 1829, 1830, 1831, 1832 et 1833. Tome sixieme, 2e. partie: Insectes*. P. Bertrand, Paris, 222 pp. + 32 pls.
- BLANCHARD E. 1847: [Insectes Coléoptères, livraison 90]. Pp. 185–222. In: d'ORBIGNY A. 1837–45 (ed.): *Voyage dans l'Amérique méridionale (Le Brésil, la République Orientale de l'Uruguay, la République Argentine, la Patagonie, la République du Chili, la République de Bolivie, la République du Pérou), exécuté pendant les années 1826, 1827, 1828, 1829, 1830, 1831, 1832 et 1833. Tome sixieme, 2e. partie: Insectes*. P. Bertrand, Paris, 222 pp. + 32 pls.
- BONDAR G. 1940: A posição do genero Himatidium na familia dos Crisomelideos e descrição de uma especie nova. *O Campo* **11**: 37–38.
- BRYANT G. E. 1942: Lista provisional de los crisomelidos (Coleoptera) de Venezuela. *Boletín de la Sociedad Venezolana de Ciencias Naturales* **8**: 195–208.
- CHABOO C. S. & STAINES C. L. 2015: Beetles (Coleoptera) of Peru: A survey of the families. Chrysomelidae: Cassidinae Gyllenhal sensu lato. *Journal of the Kansas Entomological Society* **88**: 387–398.
- CHAMPION G. C. 1907: Itinerary of Mr. G. C. Champion's travels in Central America, 1879–1883. *Entomological News* **18**: 33–44.
- DESCARPENTRIES A. & VILLIERS A. 1959: Les types de la collection M. Pic. I. Chrysomelinae et Hispinae du nouveau-monde. *Bulletin du Museum National d'Histoire Naturelle Paris* **31**: 137–154.
- FLOWERS R. W. & HANSON P. E. 2003: The diversity of the Chrysomelidae fauna in Costa Rica: Insights from a Malaise trapline. Pp. 25–51. In: FURTH D. G. (ed.): *Special Topics in Leaf Beetle Biology*. Pensoft, Series Faunistica 29, Sofia-Moscow, xii + 339 pp.
- JALINSKY J. R., WERTENBERGER R. A., RADOY T. A. & CHABOO C. S. 2014: Insects inhabiting two host plants, *Heliconia stricta* Huber (Heliconiaceae) and *Calathea lutea* Schult (Marantaceae), in southeastern Peru. *Journal of the Kansas Entomological Society* **87**: 299–311.
- McKENNA D. D. & FARRELL B. D. 2005: Molecular phylogenetics and evolution of host plant use in the Neotropical rolled leaf 'hispinae' beetle genus *Cephaloleia* (Chevrolat) (Chrysomelidae: Cassidinae). *Molecular Phylogenetics and Evolution* **37**: 117–131.
- MONRÓS F. 1945: Tres interesantes confusiones en Chrysomeloidea neotropicales. *Revista de la Sociedad Entomologica Argentina* **12**: 410–415.
- MONRÓS F. & VIANAM J. 1947: Revisión sistemática de los Hispidiae argentinos (Insecta, Coleopt. Chrysomeloid.). *Annales del Museo Argentino de Ciencias Naturales Bernardino Rivadavia* **42**: 125–324 + 27 pls.
- PIC M. 1923: Nouveautés diverses. *Mélanges Exotico-Entomologiques* **38**: 1–32.

- PIC M. 1926a: Nouveautés diverses. *Mélanges Exotico-Entomologiques* **46**: 1–32.
- PIC M. 1926b: Nouveautés diverses. *Mélanges Exotico-Entomologiques* **47**: 1–32.
- PIC M. 1934: Nouveautés diverses, notes. *Mélanges Exotico-Entomologiques* **63**: 1–36.
- SEKERKA L. 2014: Review of Imatidiini genera (Coleoptera: Chrysomelidae: Cassidinae). *Acta Entomologica Musei Nationalis Pragae* **54**: 257–314.
- SHERBORN C. D. & GRIFFIN F. J. 1934: On the dates of publication of the natural history portions of Alcide d'Orbigny's 'Voyage Amérique méridionale.' *Annals and Magazine of Natural History, Tenth Series* **13**: 130–134.
- STAINES C. L. 1996: The genus *Cephaloleia* (Coleoptera: Chrysomelidae) in Central America and the West Indies. *Revista de Biología Tropical, Special Publication* **3**: 1–73 + 14 pls.
- STAINES C. L. 2002: The new world tribes and genera of hispines (Coleoptera: Chrysomelidae: Cassidinae). *Proceedings of the Entomological Society of Washington* **104**: 721–784.
- STAINES C. L. 2009: Generic reassignment of species in the tribe Cephaloleiini Chapuis, 1875 (Coleoptera: Cassidinae). *Insecta Mundi* **107**: 1–4.
- STAINES C. L. 2013: *Aslamidium* (Neolasmidium) apertura a new species from Venezuela (Coleoptera: Chrysomelidae: Cassidinae). *Proceedings of the Entomological Society of Washington* **115**: 286–288.
- STAINES C. L. & GARCÍA-ROBLEDO C. 2014: The genus *Cephaloleia* Chevrolat, 1836 (Coleoptera, Chrysomelidae, Cassidinae). *ZooKeys* **436**: 1–355.
- UHMANN E. 1930a: Amerikanische Hispinen (Col. Chrys.). 19. Beitrag zur Kenntnis der Hispinen. *Entomologische Blätter* **26**: 33–38.
- UHMANN E. 1930b: Hispinen aus Costa Rica aus der Ausbeute des Herrn Ferd. Nevermann. 20. Beitrag zur Kenntnis der Hispinen (Col. Chrys.). *Folia Zoologica et Hydrobiologica* **1**: 209–256.
- UHMANN E. 1933: Drei neue südamerikanische Hispinen. 41. Beitrag zur Kenntnis der Hispinen (Col. Chrys.). *Folia Zoologica et Hydrobiologica* **5**: 77–81.
- UHMANN E. 1939: Hispinen des Deutschen Entomologischen Instituts, Berlin-Dahlem. III. Teil. (Coleoptera: Chrysomelidae). 82. Beitrag zur Kenntnis der Hispinen. *Arbeiten über Morphologische und Taxonomische Entomologie aus Berlin-Dahlem* **6**: 151–156.
- UHMANN E. 1940: Hispinen des Deutschen Entomologischen Instituts, Berlin-Dahlem. IV. Teil. (Coleoptera: Chrysomelidae). 89. Beitrag zur Kenntnis der Hispinen. *Arbeiten über Morphologische und Taxonomische Entomologie aus Berlin-Dahlem* **7**: 113–120.
- UHMANN E. 1957a: *Chrysomelidae: Hispinae. Hispinae Americanae*. In: HINCKS W. D. (ed.): *Coleopterorum Catalogus Supplementa, Pars 35, Fasc. 1*. Dr. W. Junk, 's-Gravenhage, vii + pp. 1–153.
- UHMANN E. 1957b: *Hispinae* aus dem Ungarischen National-Museum in Budapest. 180. Beitrag zur Kenntnis der Hispinae (Coleopt. Chrysomelidae). *Annales Historico-Naturales Musei Nationalis Hungarici, Series Nova* **8**: 301–306.
- UHMANN E. 1964: *Chrysomelidae: Hispinae. Corrigenda et Addenda*. In: STEEL W. O. (ed.): *Coleopterorum Catalogus Supplementa. Pars 35, Fasc. 3*. W. Junk, 's-Gravenhage, v + pp. 399–490.
- UHMANN E. 1968: Neue Hispinae aus Brasilien (232. Beitrag zur Kenntnis der Hispinae; Coleoptera, Chrysomelidae). *Papéis Avulsos de Zoologia* **21**: 125–130.
- WEISE J. 1905: Beschreibung einiger Hispinen. *Archiv für Naturgeschichte* **71**: 49–104.
- WEISE J. 1910: Beitrag zur Kenntnis der amerikanischen Hispinen. *Archiv für Naturgeschichte* **76**: 67–127.
- WEISE J. 1911: *Chrysomelidae: Hispinae*. In: SCHENKLING S. (ed.): *Coleopterorum Catalogus, Pars 35*. W. Junk, Berlin, 94 pp.