

***Svatactesis* gen. nov., a new genus for *Polyctesis johanidesi*
with notes on the tribe Polyctesini
(Coleoptera: Buprestidae: Polycestinae)**

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Abstract. *Svatactesis* gen. nov. is established for *Polyctesis johanidesi* Bílý, 1997 known from Turkey and Iran, and compared with closely related genera *Polyctesis* Marseul, 1865, *Bellamyina* Bílý, 1994, and *Schoutedeniastes* Burgeon, 1941. *Svatactesis johanidesi* is re-described and illustrated. Composition, taxonomic state and relations of Polyctesini Cobos, 1955 and diagnostic characters of the genera belonging to the *Polyctesis* generic-group are discussed. The *Schoutedeniastes magnifica* (Waterhouse, 1875), *S. amabilis* (Laporte & Gory, 1835) and *S. vitalisi* (Bourgoin, 1922) species-groups are distinguished within the genus *Schoutedeniastes*. Evident variability of the main diagnostic characters among species-groups cast doubts on the monophyly of this genus. First exact record of *Svatactesis johanidesi* from Iran is provided.

Key words. Coleoptera, Buprestidae, Polycestinae, Polyctesini, *Polyctesis* generic-group, *Svatactesis*, new genus, Iran, Turkey, Palaearctic Region

Introduction

BÍLÝ (1997) described *Polyctesis johanidesi* Bílý, 1997 from Turkey and indicated that in many character states and bionomy it significantly differs from another West-Palaearctic species, *P. rhois* Marseul, 1865, also occurring in Turkey. The comparison of *P. johanidesi* with other species of *Polyctesis* Marseul, 1865 and related genera attributed to the *Polyctesis* generic-group of the tribe Polyctesini Cobos, 1955 has shown that based on a set of unique character states *P. johanidesi* should be placed in a separate genus which is described below. At present the generic composition of Polyctesini is still open to debate, and taxonomic rank, content and relations of the constitutive genera need revision. Among them *Bellamyina* Bílý, 1994 and *Schoutedeniastes* Burgeon, 1941 are the most debatable genera (COBOS 1955; HOŁYŃSKI 1993, 2003; BELLAMY 2003, 2008; VOLKOVITSH 2008).

Material and methods

Photographs of the habitus were taken using a Canon D-550 digital camera with a Canon MP-E65 mm f/2.8 1–5× macro lens (NMPC) and an Olympus SZ-CTV dissecting microscope with mounted Olympus-Camedia 3030 Zoom camera (ZIN); photographs of the morphological structures were taken using a Leica MZ-9.5 stereomicroscope with mounted Leica DFC-290 camera and Bresser-Biolux light microscope with integrated imaging system (ZIN). Measurements were taken using eyepiece micrometer in an MBS-9 stereomicroscope.

The examined specimens are deposited in the following collections:

- CDFA California Department of Food and Agriculture (C. L. Bellamy collection), Sacramento, U.S.A.;
MJCG Martin Johanides collection, Prague, Czech Republic;
MNHN Muséum national d'Histoire naturelle, Paris, France;
NMPC National Museum, Prague, Czech Republic;
VKCB Vítězslav Kubáň collection, Šlapanice, Czech Republic;
VCCP Vladimír Čelikovský collection, Prague, Czech Republic;
ZIN Zoological Institute, Russian Academy of Sciences, St. Petersburg, Russia.

Taxonomy

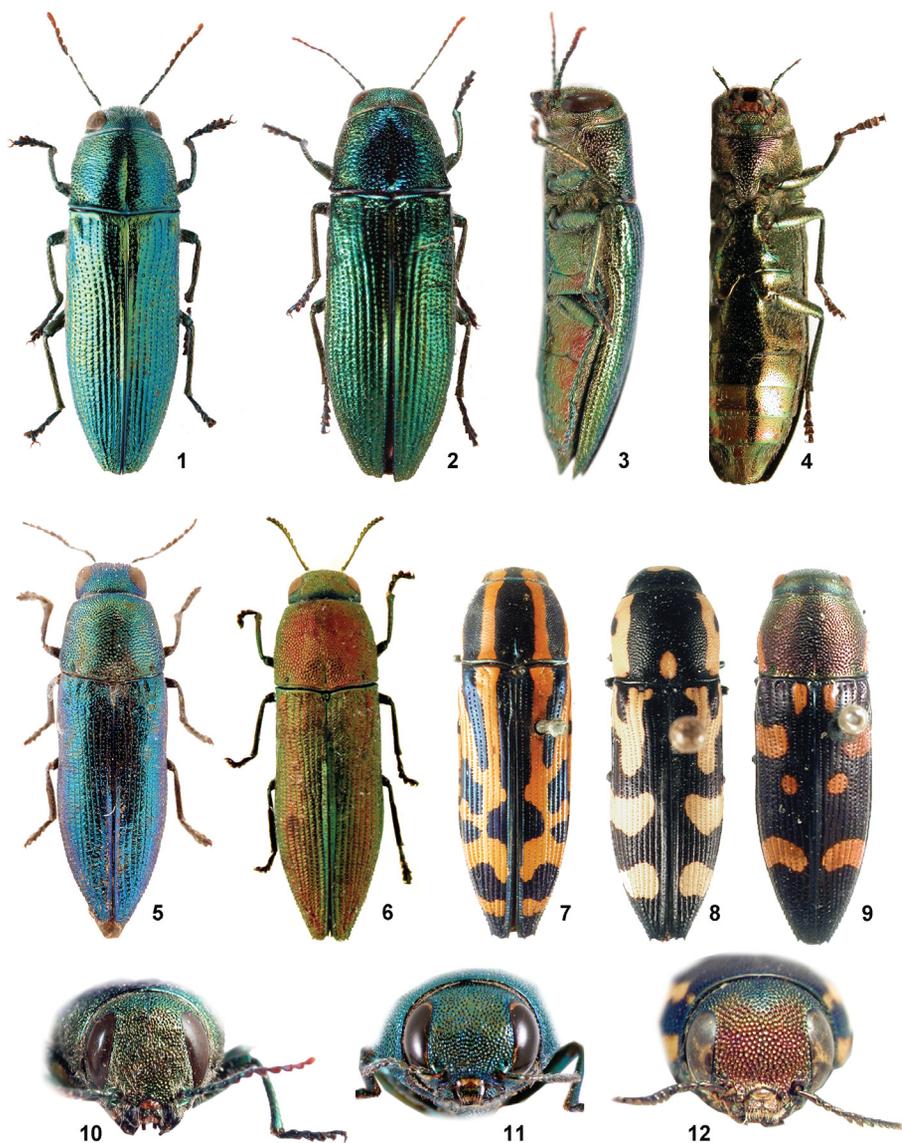
Svatactesis gen. nov.

Type species. *Polyctesis johanidesi* Bílý, 1997, here designated.

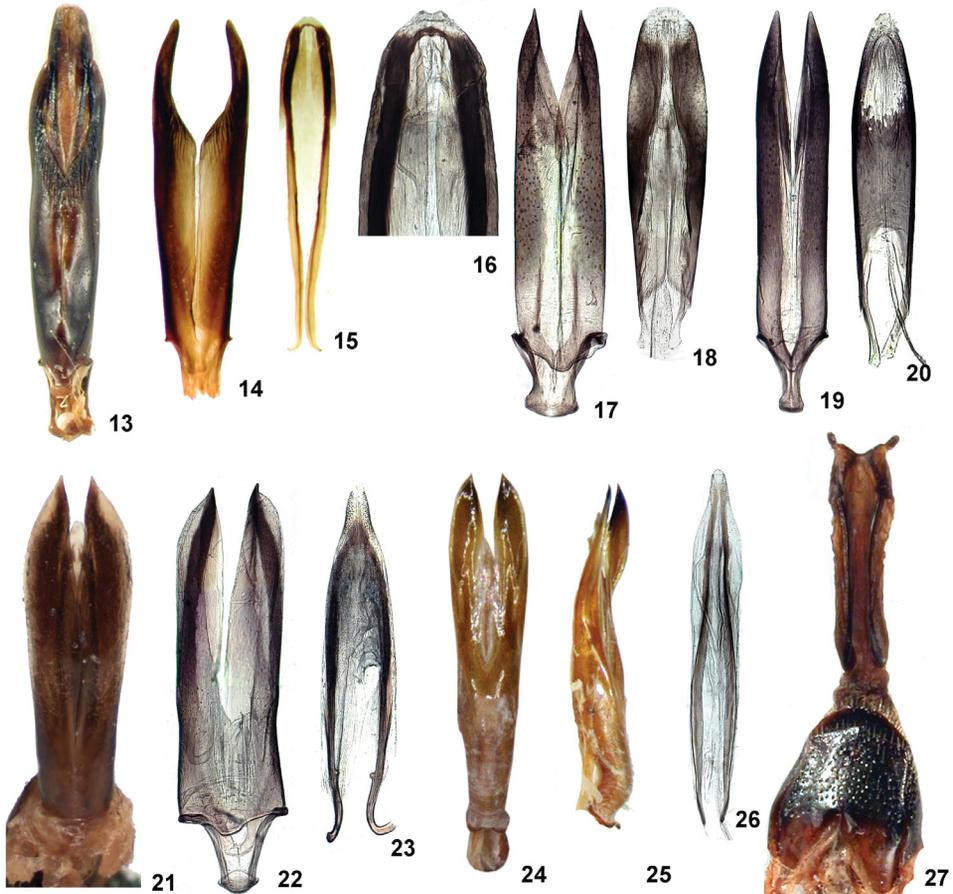
Description. Medium sized, metallic, iridescent, golden-green or bluish-green dorsally, elytra with indistinct golden-bronze marginal stripe along entire length; frons distinctly narrowing towards vertex, clypeus with narrow deep emargination anteriorly, antennal sockets open, antennae bicoloured; pronotal disk with deep medial depression, covered with very coarse transversely rugose sculpture, posterior angles of pronotum sharply protruding outward; scutellum absent; subhumeral lobe of elytra narrow, not covering entire metepisternum; lateral margin nearly smooth in apical half of elytra, apices obtusely bidentate; 2nd stria of elytra very short reaching only 1/4–1/5 of elytral length; pro- and mesotibiae dimorphic; penis with short (about 1/3 of entire length), sclerotized apical part and very long (about 2/3 of entire length) basal apophyses (see also under *S. johanidesi*).

Differential diagnosis (Table 1). *Svatactesis* gen. nov. is easily distinguished from other members of *Polyctesis* generic-group by frons distinctly narrowing towards vertex, pronotum with deep medial depression and strongly protruding outward posterior angles, absence of scutellum, unique penis structure, and larval host plant (see below). It shares some character states with *Polyctesis* and *Bellamyina* (metallic iridescent coloration, subhumeral lobe of elytra narrow) on the one hand, and with *Schoutedeniastes* (clypeus with deep and narrow anterior emargination, antennal sockets open, and some other characters within the species-groups) on the other hand.

Etymology. Generic name is derived from *Polyctesis* (feminine: BELLAMY 2008). It is dedicated to Svatopluk (Svát'a for friends) Bílý in honor of our 40-year friendship and collaboration.



Figs 1–12. 1–9 – habitus. 1–4 – *Svatactesis johanidesi* (Bilý, 1997). 1 – holotype, ♂ (NMPC), 13.6 mm. 2–4 – ♀, Turkey (Aydinlar) (VKCB), 14.3 mm: 2 – dorsal view, 3 – lateral view, 4 – ventral view. 5 – *Polycyctes rhois* Marseul, 1865, ♂, Turkey (Erzincan) (ZIN), 11.9 mm. 6 – *Bellamyina hunanensis* (Peng, 1992), ♀, China (Shaanxi) (ZIN), 12.0 mm. 7 – *Schoutedeniastes magnifica* (Waterhouse, 1875), syntype, ♀, “Limpopo” (MNHN), 15.2 mm. 8 – *S. amabilis* (Laporte & Gory, 1835), ♀, Sri Lanka (MNHN), 11.2 mm. 9 – *S. vitalisi* (Bourgoin, 1922), ♀, syntype, Laos (Muong You) (MNHN), 12.3 mm. 10–12 – head, frontal view. 10 – *Svatactesis johanidesi*, ♀ (VKCB). 11 – *Polycyctes rhois*, ♀, Turkey (ZIN). 12 – *Schoutedeniastes amabilis* (Laporte & Gory, 1835), ♂, India (Tamil Nadu) (NMPC). Photo: 1–4, 10 – V. Kubán; 5–9, 11, 12 – M. Volkovitsh.



Figs 13–27. 13–16 – male genitalia. 13–16 – *Svatactesis johanidesi* (Bilý, 1997), dorsal view: 13 – aedeagus, holotype; 14–16 – paratype, Turkey (Aslanli) (NMPC): 14 – tegmen; 15 – penis; 16 – apex of penis. 17–18 – *Polyctesis rhois* Marseul, 1865, Turkey (Erzincan), dorsal view. 17 – tegmen; 18 – penis. 19–20 – *Bellamyina hunanensis* (Peng, 1992), China (Shaanxi), dorsal view: 19 – tegmen; 20 – penis. 21 – *Schoutedeniastes magnifica* (Waterhouse, 1875), Zimbabwe (CDFA), aedeagus, ventral view. 22–23 – *S. okhurai* (Akiyama & Ohmomo, 1992), Thailand (ZIN), dorsal view: 22 – tegmen; 23 – penis. 24–26 – *S. vitalisi* (Bourgoin, 1922), Thailand (NMPC): 24–25 – tegmen: 24 – dorsal view; 25 – lateral view. 26 – penis, dorsal view. 27 – *Svatactesis johanidesi*, ♀, Turkey (Aydinlar) (VKCB), ovipositor and pregenital abdominal segments, ventral view. Photo: 13, 27 – V. Kubáň; 14–26 – M. Volkovitsh.

Svatactesis johanidesi (Bilý, 1997)

(Figs 1–4, 10, 13–16, 27)

Polyctesis johanidesi Bilý, 1997: 15 (original description); VOLKOVITSH (2006): 341 (catalogue); BELLAMY (2008): 361 (catalogue); GHAHARI et al. (2015): 32 (catalogue).

Type locality. Turkey, Mersin Province, Erdemli to Arslanlı.

Type material (cited after Bilý 1997). HOLOTYPE: ♂, TURKEY: MERSIN PROVINCE: Erdemli-Aslanli, 900 m a.s.l., 6.–14.vi.1996, M. Johanides leg. (NMPC). ALLOTYPE: ♀, same data (NMPC). PARATYPES: same data, 7 ♂♂ 5 ♀♀ (NMPC, MJCG). TURKEY: MERSIN PROVINCE: Güzeloluk (Erdemli), 800–1200 m a.s.l., 5.–14.vi.1996, 4 ♂♂ 1 ♀, V. Čelikovský leg. (NMPC, VCCP).

Additional material examined. **TURKEY:** MERSIN PROVINCE: Aydinlar (NW of Erdemli), 23.–24.vi.1998, 1 ♀, J. Chalupek leg. (VKCB). **IRAN:** LORESTAN PROVINCE: 10 km SE of Bavineh, 1100 m a.s.l., 33°36'08"N, 47°11'59"E, 16.–17.x.1998, 1 ♀, P. Kabátek leg. (NMPC).

Redescription. Body (Figs 1–4) elongate, 3.27 (3.10–3.62) times as long as pronotum width at base, slightly convex, without dorsal curvature; dorsally golden-green, occasionally bluish-green, elytra with poorly marked golden-bronze marginal stripe along entire length, ventrally golden-bronze to golden-red; antennal segments VI–XI fulvous, proximal segments and legs golden-green; body dorsally covered with short, semi-erect, white setae, ventrally with longer recumbent setae. Total length 14.2 (12.2–16.1) mm, width 4.4 (3.7–5.2) mm (n = 8: 4 ♂♂ 4 ♀♀).

Head (Fig. 10) relatively narrow, moderately convex when seen from above (Figs 1–2); frons flattened, without medial line or depression, with nearly straight, markedly convergent sides. Vertex with fine medial line, narrow, 1.65 (1.40–2.00) times as wide as transverse diameter of eye and 0.74 (0.70–0.81) times as wide as frons above antennal sockets. Eyes large, moderately convex, slightly or not protruding beyond head outline. Clypeus: lateral branches nearly completely reduced, not enclosing antennal sockets from below (sockets open); with narrow, deep, arcuate medial emargination anteriorly. Anteclypeus exposed, with medial groove. Frons with coarse reticulate-rugose sculpture of small, deep, irregular, umbilicate punctures without inner granules and poorly marked micropunctures; intervals narrower than diameter of puncture, forming slightly elevated transverse rugosities; covered with relatively long, semi-erect white setae. Antennae very long, in male 2.22 (2.09–2.31), in female 2.20 (2.05–2.32) times as long as height of eye; enlarged from antennomere V and bearing large sensory fossae ventrally from antennomere IV; antennomeres IV–X obtusely-serrate, much longer than wide; antennomere XI irregularly oval, 1.5–2.0 times as long as wide; antennomeres VI–XI partly or completely fulvous in both sexes.

Pronotum (Figs 1–3) more or less bell-shaped with acute posterior angles protruding outward, slightly transverse, 1.43 (1.35–1.48) times as wide at base as long, widest at base; pronotal sides S-shaped. Anterior margin feebly bisinuate, projecting forward, bordered with fine groove; basal margin slightly angularly protruding medially, nearly straight. Lateral carina fine, strongly incurved, S-shaped, entire, nearly reaching anterior angles. Pronotal surface flattened, with large, deep medial depression; prescutellar fossa poorly marked at base of medial depression, latero-basal depressions punctiform. Pronotal sides with ocellate sculpture of deep punctures with inconspicuous inner structure and elevated intervals; toward disc intervals merge, forming transverse rugae, disc covered with very coarse punctato-rugose sculpture. Sides with relatively long, recumbent, white setae; disc with inconspicuous, sparse, short setae. Anterior prosternal margin (Fig. 4) straight, bordered with well marked, fine groove; prosternum weakly convex, covered with coarse punctato-rugose sculpture; prosternal process wide, slightly narrowing apically, covered with dense simple punctures. Hypomerion bearing dense, nearly alveolate sculpture. Scutellum entirely absent (Figs 1, 2).

Elytra (Figs 1–3) moderately elongate, 2.35 (2.22–2.44) times as long as wide at base, weakly convex; sides not widening at humeri, subparallel or slightly diverging toward posterior 1/3, then gently arcuately converging to apices; apices obtusely bidentate, with poorly marked sutural and angular marginal denticles, the later sometimes blunt; lateral margin slightly deflexed, epipleura poorly separated by indistinct carina; subhumeral lobe not cov-

ering metepisternum, bearing small denticle posteriorly; epipleural serrations inconspicuous, margin nearly smooth in posterior half. Strial punctures round or slightly elongate, separated in anterior half; in posterior half punctures merging and striae slightly sulcate; 2nd stria short, reaching about 1/4–1/5 of elytral length. Intervals subequal, weakly convex or flattened, on disc about five times as wide as diameter of strial punctures; discal intervals with very fine, sparse uniseriate punctures, lateral intervals finely transversely rugulose; covered with fine, curved, uniseriate white setae, approximately as long as half of interval width. Elytra golden green, occasionally bluish-green, with indistinct golden-bronze marginal stripe along entire length, sometimes extending to pronotal sides.

Legs (Figs 1–4). Femora and tibiae ventrally golden-green with bronzy sheen, tibiae dorsally copper-green, tarsi bluish; metacoxal plates (Figs 3–4) with subparallel margins, posterior margin nearly straight, slightly emarginate laterally, without lateral tooth. Tibiae feebly widened toward apices; in male pro- and mesotibiae strongly curved (Fig. 1), in female protibiae slightly curved, mesotibiae straight (Fig. 2); metatibiae bearing comb of dense yellowish setae externally. Tarsomere I longest, equal to II and III combined; tarsomeres II–IV subequal, short; tarsomere IV with medial notch at anterior margin; tarsomere V relatively short, equal to III and IV combined, flattened and distinctly expanded toward apex; tarsal pads well developed on tarsomeres I–IV, on tarsomere I as long as tarsomere itself. Tarsal claws long, curved, simple, swollen at base.

Abdomen (Figs 3–4) golden-green with coppery sheen medially changing to golden-red laterally; covered with dense simple punctures (intervals equal to 2–3 diameters of a puncture laterally and more than 3 diameters medially) and rather long semi-erect white setae which are denser on sides. Suture between ventrites I–II well marked, nearly straight. Anal ventrite obtusely rounded apically in both sexes.

Male. Aedeagus as in Figs 13–16. Parameres dorsally with distinct longitudinal rugosity not extending to their apices (Figs 13–14). Penis (Figs 15–16) with very short (about 1/3 of entire length), sclerotized, obtuse apical part and very long (about 2/3 of entire length), nearly straight basal apophyses with curved apices.

Female. Ovipositor (Fig. 27) of tubular type, relatively short, approximately 2.5 times as long as expanded apical part, with angularly emarginate apex and styli widely separated from each other.

Differential diagnosis. See under diagnosis of *Svatactesis* gen. nov. and Table 1. Additionally, *S. johanidesi* differs from a single West-Palaearctic species of Polyctesini, *Polyctesis rhois* (Fig. 5), as follows (see also BÍLÝ 1997): dorsal and ventral coloration different (in *P. rhois* nearly unicolorous); pronotum and elytra bearing distinct setae as long as half of width of elytral intervals (in *P. rhois* inconspicuous, nearly as long as diameter of punctures); sides of frons distinctly converging towards vertex (in *P. rhois* almost parallel sided or slightly converging, 0.94 (0.90–1.00) times as wide as frons above antennal sockets (Fig. 11)); antennae very long, antennomeres VI–XI fulvous (in *P. rhois* antennae much shorter, in male 1.63 (1.43–1.70), in female 1.32–1.66 times as long as vertical diameter of eye, unicolorous); pronotum more strongly transverse, rugosities very coarse, lateral carina complete (in *P. rhois* pronotum 1.17 (1.13–1.26) times as wide at base as long, transverse rugosities fine, lateral carina incomplete); aedeagus elongate, narrow (Figs 13, 14), with longitudinal dorsal rugosities not extending to parameral apices, penis with very short sclerotised apical part and very long basal apophyses

Table 1. Comparison of the main taxonomic characters among the genera of *Polyctesis* generic-group

	<i>Bellamyina</i>	<i>Polyctesis</i>	<i>Svatactesis</i>		
	Bily, 1994	Marseul, 1865	gen. nov.	<i>magnifica</i>	<i>amabilis</i>
				species-group	species-group
				<i>amabilis</i>	<i>vitalisi</i>
Coloration	Metallic, iridescent (Fig. 6)	Metallic, nearly unicolorous (Fig. 5)	Metallic, elytra with marginal stripe, ventrally iridescent (Figs 1–4)	Multicolorous, with pigmented markings (Fig. 7)	Multicolorous, with pigmented markings (Fig. 8)
Frons, shape	Margins subparallel or slightly convergent (Fig. 11)	Margins subparallel or slightly convergent (Fig. 11)	Margins distinctly convergent (Fig. 10)	Margins subparallel or slightly convergent (Fig. 12)	Margins subparallel or slightly convergent (Fig. 12)
Clypeus, anterior emargination	Shallow	Shallow (Fig. 11)	Deep (Fig. 10)	Deep (Fig. 12)	Deep
Antennal sockets	Closed	Closed (Fig. 11)	Open (Fig. 10)	Open (Fig. 12)	Open
Pronotum, sculpture	Uniformly alveolate (Fig. 6)	Ocellate, disc transversely rugose (Fig. 5)	Ocellate, disc coarsely transversely rugose (Figs 1, 2)	Alveolate (Fig. 7)	Ocellate (Fig. 8)
Pronotum, medial depression	Line, mainly at base (Fig. 6)	Line or indistinct depression (Fig. 5)	Distinct deep depression (Figs 1, 2)	Line, mainly at base (Fig. 7)	Absent/line at base (Fig. 9)
Pronotum, latero-basal corners	Not protruding, obtuse (Fig. 6)	Not protruding, obtuse (Fig. 5)	Strongly protruding, acute (Figs 1, 2)	Not protruding, obtuse (Fig. 7)	Not protruding, obtuse (Fig. 9)
Scutellum	Present (Fig. 6)	Present (Fig. 5)	Absent (Figs 1, 2)	Present (Fig. 7)	Present (Fig. 9)
Elytra: 2nd stria	Long, about 1/2 of elytral length (Fig. 6)	Longer than 1/2 of elytral length (Fig. 5)	Short, 1/4–1/5 of elytral length (Figs 1–2)	Short, 1/4–1/5 of elytral length (Fig. 7)	Longer than 1/2 of elytral length (Fig. 9)
Elytra, lateral margin in posterior half	Serrate (Fig. 6)	Serrate (Fig. 5)	Nearly smooth (Figs 1–4)	Serrate (Fig. 7)	Serrate/nearly smooth (Fig. 9)
Elytra, subhumeral lobe	Narrow, not covering metepisternum, with small tooth posteriorly	Narrow, not covering metepisternum, with small tooth posteriorly	Narrow, not covering metepisternum, with small tooth posteriorly (Fig. 3)	Broad, nearly covering metepisternum, without tooth posteriorly	Broad, nearly covering metepisternum, with small tooth posteriorly
Fore and middle tibiae, male	Straight	Straight (Fig. 5)	Distinctly incurved (Fig. 1)	Nearly straight	Distinctly incurved or with large teeth apically
Distribution	Eastern Palaearctic	Palaearctic, South-East Asia	Western Palaearctic	Africa	South-East Asia, Eastern Palaearctic
Host plants	<i>Cotinus</i>	<i>Rhus</i>	Unknown	Unknown	<i>Acacia</i>

(Fig. 15) (in *P. rhois* aedeagus much wider and shorter (Fig. 17), dorsal rugosity extending to parameral apices (see BÍLÝ 1997: Fig. 5), penis with very long sclerotized apical part and very short basal apophyses (Fig. 18); larvae develop in *Quercus* spp. (Fagaceae) (larvae of *P. rhois* develop in *Rhus coriaria* L. (Anacardiaceae)).

Ecological information. Larvae of *Svatactesis johanidesi* develop in dead branches of *Quercus* spp. (Fagaceae) (BÍLÝ 1997) in contrast to the known larvae of *Polyctesis* (*P. rhois* and *P. strandi* Obenberger, 1934) and *Bellamyina* (*B. hunanensis* (Peng, 1992)) whose larvae develop in the species of Anacardiaceae (BÍLÝ 1997; VOLKOVITSH 2004; E. Kučera, pers. comm.). The only known larval host plant of *Schoutedeniastes* is *Acacia gageana* Craib (Fabaceae) (1 spec. of *S. apicata* (Waterhouse, 1882) from Dehradun, Uttarkhand, India (MNHN)). Adults of *S. birmanica* (Théry, 1947) have been collected on the leaves of unidentified tree species of Fabaceae in Laos (V. Kubáň, personal communication).

Distribution. Turkey (Mersin) (BÍLÝ 1997), Iran (Lorestan) (GHAHARI et al. 2015, this paper).

Discussion

Up until now the generic composition of the tribe Polyctesini remains fairly debatable. COBOS (1955) established a new tribe within subfamily Polycestinae Lacordaire, 1857 for two genera previously belonging to Ptosimini Kerremans, 1892: *Polyctesis* (including *Schoutedeniastes*) and *Chrysophana* LeConte, 1860. Later, COBOS (1980) attributed to Polyctesini the genera *Xenopsis* Saunders, 1867 and newly established *Paraxenopsis* Cobos, 1980. This generic composition was adopted by BELLAMY (1985) with addition of *Beerellus* Nelson, 1982 closely related to *Chrysophana*. HOLYŃSKI (1993) suggested a completely different concept: subtribe Polyctesina *sensu novo* which comprised only *Polyctesis* and *Schoutedeniastes* (as full genus) was attributed to the tribe Thrincopygini LeConte, 1861 (subfamily Buprestinae Leach, 1815) while *Chrysophana* and *Beerellus* were transferred to the subtribe Bubastina Obenberger, 1920 belonging to the tribe Anthaxiini Gory & Laporte, 1839. VOLKOVITSH (2001) based on the comparative study of antennal sensory organs established within Polycestoid complex (= Polycestinae) a separate Polyctesiod lineage which comprised the tribes Thrincopygini (*Thrincopyge* LeConte, 1858), Polyctesini (*Polyctesis*, *Bellamyina*, *Schoutedeniastes*, *Paraxenopsis*), and informal *Chrysophana* generic-group (*Chrysophana*, *Beerellus*) (only genera with studied antennal structures were considered). HOLYŃSKI (2003) re-examined Polyctesina *sensu* HOLYŃSKI (1993) and separated two generic-groups: *Polyctesis*-group (*Polyctesis* including *Bellamyina* as its subgenus and *Schoutedeniastes*) and *Xenopsis*-group (*Xenopsis*, *Paraxenopsis*, *Sommaia* Toyama, 1985, and *Kurosawaxia* Descarpentries, 1986). A similar generic composition of Polyctesini (generic level of *Bellamyina* resurrected) was accepted by BELLAMY (2003, 2008). VOLKOVITSH (2008) transferred genera of *Xenopsis*-group to the newly established subtribe Xenopseina Volkovitsh, 2008 (as Xenopsina) within the tribe Polycestini Lacordaire, 1857, synonymized Polyctesiod lineage under Polycestioid lineage, and indicated, that *Chrysophana* generic-group most probably should be returned to Polyctesini (see also VOLKOVITSH 2006). At present, the taxonomic structure of the tribe Polyctesini is as follows:

***Chrysophana* generic-group Volkovitsh, 2001**

Chrysophana LeConte, 1860; type species *Ancylocheira placida* LeConte, 1854, by monotypy; two species in Nearctic, one species in Palaearctic (Himalaya) (BELLAMY 2008).

Beerellus Nelson, 1982; type species *Beerelus taxodii* Nelson, 1982, by monotypy; one species in Nearctic (BELLAMY 2008).

***Polyctesis* generic-group Holyński, 2003**

Bellamyina Bílý, 1994 (Figs 6, 19, 20); type species *Bellamyina cylindrica* Bílý, 1994 (= *Polyctesis hunanensis* Peng, 1992), by original designation; one species in East Palaearctic (VOLKOVITSH 2006, BELLAMY 2008).

Polyctesis Marseul, 1865 (Figs 5, 11, 17, 18); type species *Polyctesis rhois* Marseul, 1865, by monotypy; one species in West Palaearctic (East Mediterranean), three species in East Palaearctic and Oriental Region.

Schoutedeniastes Burgeon, 1941 (substitute name for *Schoutedenia* Obenberger, 1924, nom. preocc.) (Figs 7–9, 12, 21–26); type species *Schoutedenia kerremansi* Obenberger, 1924 (= *Ptosima magnifica* Waterhouse, 1875), by original designation; one species in Afrotropical Region, nine species in Oriental Region and East Palaearctic.

Svatactesis gen. nov. (Figs 1–4, 10, 13–16, 27); type species: *Polyctesis johannidesi* Bílý, 1997, by original designation; one species in West Palaearctic.

Formally, *Chrysophana* and *Polyctesis* generic-groups can be treated as distinct subtribes but I believe that their establishment prior to a taxonomic revision of the tribe Polyctesini is inappropriate, given the uncertainty regarding taxonomic level and composition of some taxa (e.g. *Bellamyina*, *Schoutedeniastes*).

HOLYŃSKI (1993, 2003) indicated that the main distinguishing characters of *Schoutedeniastes* and *Polyctesis* are coloration pattern and length of the 2nd elytral stria. Re-examination of the *Schoutedeniastes* species has shown that the length of the 2nd stria varies within the genus (Table 1) which allows the recognition of two groups of species: the *S. magnifica/amabilis* group (2nd stria short) and the *S. vitalisi* group (2nd stria long). *Schoutedeniastes magnifica* is the only Afrotropical species of this genus slightly differing from the species attributed to the *S. amabilis* group in some character states as indicated in Table 1; for this reason it is placed in separate *S. magnifica* species-group. There are also distinct differences in the male genital structures between the species of *S. amabilis* and *S. vitalisi* species-groups (Figs 22–26) but the internal structure of aedeagus of *S. magnifica* was not studied, externally it is similar to those of *S. amabilis* species-group (Figs 21–22). The species-group composition of the genus *Schoutedeniastes* is as follows:

***Schoutedeniastes* Burgeon, 1941**

S. magnifica species-group (Figs 7, 21): *S. magnifica* (Waterhouse, 1875) (= *S. kerremansi* (Obenberger, 1924)); Afrotropical Region.

- S. amabilis* species-group** (Figs 8, 12, 22–23): *S. amabilis* (Laporte & Gory, 1835), *S. apicata* (Waterhouse, 1882), *S. birmanica* (Thery, 1947) (= *S. rondoni* (Baudon, 1962)), *S. duaulti* (Baudon, 1962), *S. hieroglyphica* (Théry, 1904), *S. okhurai* (Akiyama & Ohmomo, 1992), ?*S. hatai* (Ohmomo & Akiyama, 1994) (not studied); Oriental Region and East Palaearctic.
- S. vitalisi* species-group** (Figs 9, 24–26): *S. igorrota* (Heller, 1891) (with subspecies *aenea* Hoscheck, 1931), *S. vitalisi* (Bourgoin, 1922) (= *S. consobrina* (Bourgoin, 1922; *S. vitalisi* (Obenberger, 1924))); Oriental Region and East Palaearctic.

The differences in some character states (Table 1) between these species-groups cast doubts on the monophyly of *Schoutedeniastes*. The only reliable characters to distinguish this genus from *Polyctesis* are the colour pattern, wider subhumeral lobe of elytra, nearly covering metepisternum, and the structure of clypeus (shape of anterior emargination, degree of reduction of the lateral branches) and, correspondingly, antennal sockets (open vs. closed). The length of the 2nd elytral stria of the species of *S. magnifica* and *S. amabilis* species-groups (2nd stria short) are similar to *Svatactesis* gen. nov. while the species of *S. vitalisi* species-group (2nd stria long) are similar to *Polyctesis* and *Bellamyina*. Interestingly, the penis structure of *Schoutedeniastes vitalisi* (Fig. 26) is rather similar to those of *Xenopsis violaceocyanea* Volkovitsh, 2008 and *X. kubani* Volkovitsh, 2008 (Polycestini: Xenopseina) (see VOLKOVITSH 2008, Figs 78, 80). In this way, *S. vitalisi* species-group of *Schoutedeniastes* may be a link between Polyctesini and Polycestini which in turn gives rise to doubts regarding the tribal level of Polyctesini. A detailed taxonomic revision of all the taxa attributed to Polyctesini is needed to clarify its generic composition and relations to Polycestini comprising the subtribes Xenopseina Volkovitsh, 2008 and Polycestina Lacordaire, 1857 (VOLKOVITSH 2008).

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